

Citation for published version:

Potter, B, Watt, JM & Baszczynski, O 2019, 'Synthesis of Terminal Ribose Analogues of Adenosine 5-Diphosphate Ribose as Probes for the Transient Receptor Potential Cation Channel TRPM2', *Journal of Organic Chemistry*, vol. 84, no. 10, pp. 6143-6157. <https://doi.org/10.1021/acs.joc.9b00338>

DOI:

[10.1021/acs.joc.9b00338](https://doi.org/10.1021/acs.joc.9b00338)

Publication date:

2019

Document Version

Peer reviewed version

[Link to publication](https://doi.org/10.1021/acs.joc.9b00338)

This document is the Accepted Manuscript version of a Published Work that appeared in final form in The Journal of Organic Chemistry, copyright © American Chemical Society after peer review and technical editing by the publisher. To access the final edited and published work see <https://pubs.acs.org/doi/10.1021/acs.joc.9b00338>

University of Bath

Alternative formats

If you require this document in an alternative format, please contact:
openaccess@bath.ac.uk

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Synthesis of terminal ribose analogues of adenosine 5'-diphosphate ribose (ADPR) as probes for the Transient Receptor Potential (TRP) cation channel TRPM2

Ondřej Baszczyński^{§2‡}, Joanna M. Watt^{§1,2}, Monika D. Rozewitz³, Andreas H. Guse³, Ralf Fliegert^{3,¶} and Barry V. L. Potter^{1,2,¶*}

¹Medicinal Chemistry & Drug Discovery, Department of Pharmacology, University of Oxford, Mansfield Road, Oxford, OX1 3QT, UK

²Wolfson Laboratory of Medicinal Chemistry, Department of Pharmacy and Pharmacology, University of Bath, Bath, BA2 7AY, UK

³The Calcium Signalling Group, Department of Biochemistry and Molecular Cell Biology, University Medical Center Hamburg-Eppendorf, Martinistrasse 52, 20246 Hamburg, Germany

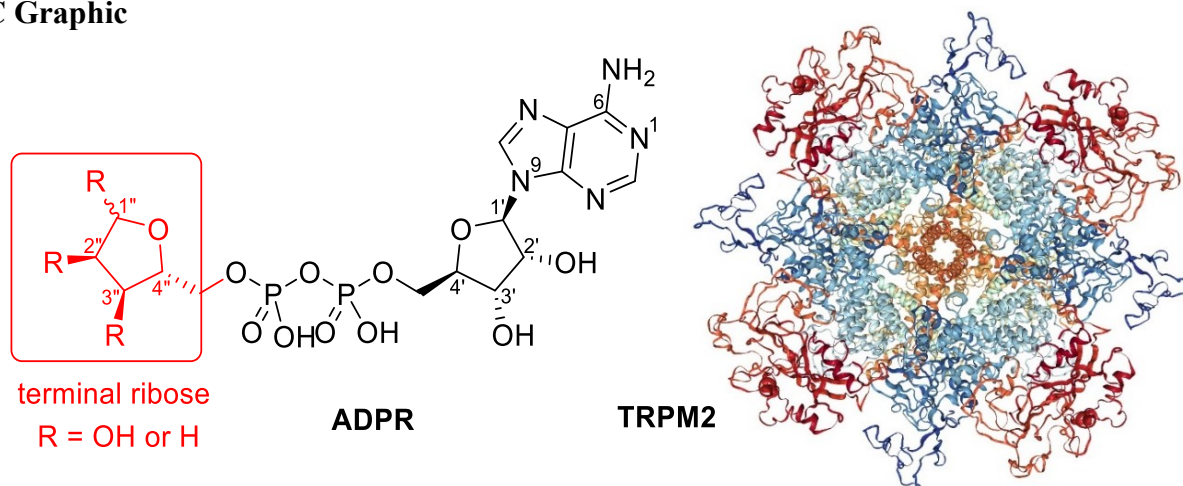
***Corresponding author** Email: barry.potter@pharm.ox.ac.uk

***Present address:** Department of Organic Chemistry, Faculty of Science, Charles University, Hlavova 2030/8, 128 43 Prague 2, Czech Republic.

§ = Equal contribution, ¶ = Equal contribution

Keywords: TRPM2, ADPR, calcium, NUDT9, pyrophosphate

TOC Graphic



Abstract

TRPM2 (transient receptor potential cation channel, subfamily M, member 2) is a non-selective cation channel involved in the response to oxidative stress and in inflammation. Its role in autoimmune and neurodegenerative diseases makes it an attractive pharmacological target. Binding of the nucleotide adenosine 5'-diphosphate ribose (ADPR) to the cytosolic NUDT9 homology (NUDT9H) domain activates the channel. A detailed understanding of how ADPR interacts with the TRPM2 ligand binding domain is lacking, hampering the rational design of modulators, but the terminal ribose of ADPR is known to be essential for activation. To study its role in more detail we designed synthetic routes to novel analogues of ADPR and 2'-deoxy-ADPR that were modified only by removal of a single hydroxyl group from the terminal ribose. The ADPR analogues were obtained by coupling nucleoside phosphorimidazolides to deoxysugar phosphates. The corresponding C2''-based analogues proved to be unstable. The C1''- and C3''-ADPR analogues were evaluated electrophysiologically by patch-clamp in TRPM2-expressing HEK293 cells. In addition, a compound with all hydroxyl groups of the terminal ribose blocked as its 1''- α -methylfuranoside-2'', 3''-isopropylidene derivative was evaluated. Removal of either C1'' or C3'' hydroxyl groups from ADPR resulted in loss of agonist activity. Both these modifications, and blocking all three hydroxyl groups resulted in ADPR antagonists. Our results demonstrate the critical role of these hydroxyl groups in channel activation.

Introduction

The non-selective cation channel TRPM2 (transient receptor potential cation channel, subfamily M, member 2) is activated in a Ca^{2+} -dependent manner after binding of the nucleotide ADP-ribose (ADPR) to its cytosolic C-terminal NUDT9H domain – that shares homology with a mitochondrial nucleotide pyrophosphatase NUDT9.¹ While earlier studies indicated that the NUDT9H domain also has a low pyrophosphatase activity, hydrolyzing ADPR to AMP and ribose 5-phosphate (R5P),² a recent study showed that the Nudix box motif in TRPM2 does not support catalysis and the production of AMP might have been due to spontaneous hydrolysis of ADPR at alkaline pH.³

Reactive oxygen species (ROS) and genotoxic stress can result in release of ADPR from the nucleus due to the activation of the poly(ADP-ribose) polymerase-1 and poly(ADP-ribose) glycohydrolase pathway.^{4,5} The ADPR so generated can then activate TRPM2, resulting in prolonged Ca^{2+} -entry, mitochondrial Ca^{2+} -overload and apoptosis,⁶ thereby contributing to cell damage in post ischemic reperfusion injury during myocardial infarction⁷ and stroke.⁸

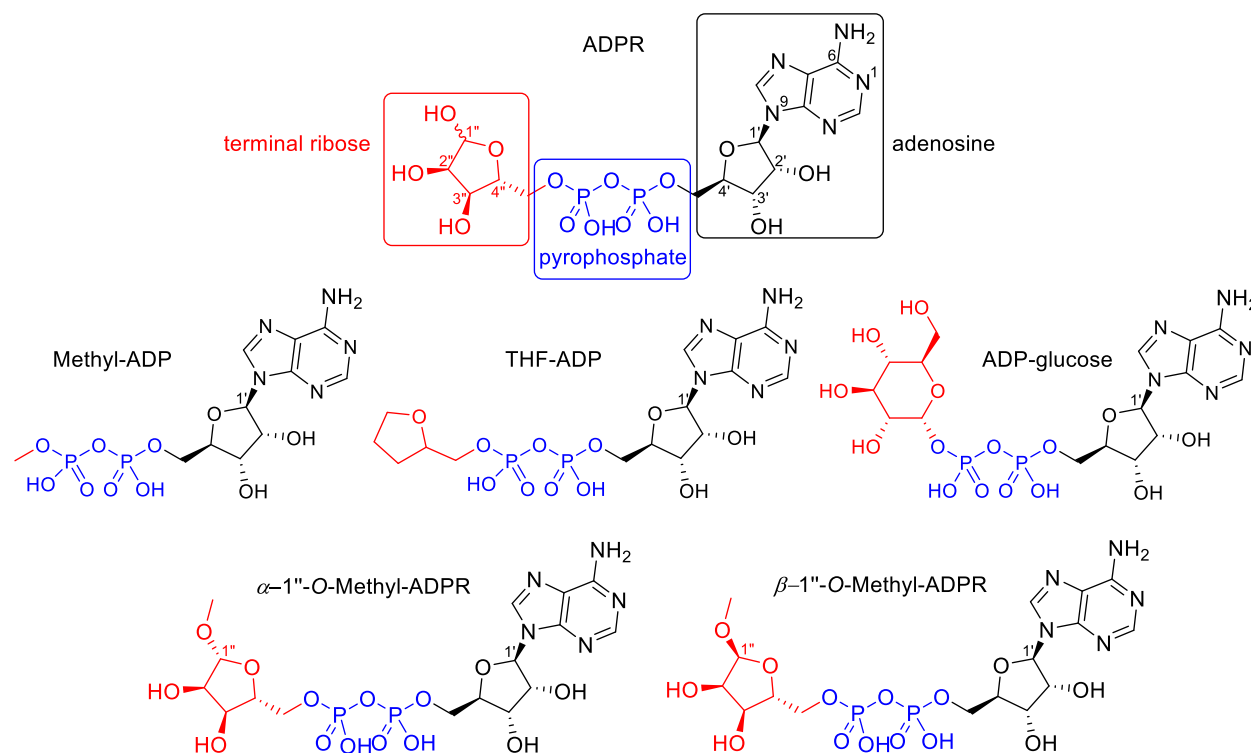
Besides this role in cell death, TRPM2 also participates in physiological processes like inflammation.⁹ In neutrophil granulocytes and dendritic cells TRPM2 contributes to chemotaxis.^{10–12} The chemotaxis of murine neutrophils in response to fMLP is independent of PARP-1 but can be inhibited by 8-Br-ADPR, a compound that inhibits activation of TRPM2 by ADPR, and by knock-out of CD38, a glycohydrolase that produces ADPR from NAD.¹⁰ In macrophages and monocytes TRPM2 is involved in secretion of chemokines and cytokines in response to ROS and pro-inflammatory cytokines,^{13,14} whereas in effector T cells it plays a role in proliferation and secretion of pro-inflammatory cytokines.¹⁵ Inhibition of TRPM2 has been shown to reduce tissue damage after stroke by preventing invasion of neutrophils¹⁶ and knock-out of TRPM2 ameliorates the symptoms of experimentally induced autoimmune encephalomyelitis, a model for multiple sclerosis.¹⁵ TRPM2 is thus an attractive pharmacological target for the treatment of neurodegenerative and autoimmune diseases.^{17,18}

Further investigations into the role of TRPM2 in physiology and pathophysiology require specific modulators of channel function and their rational design will benefit from a structure of the binding site and a better understanding of its structure activity relationship (SAR). Recently, TRPM2 structures from three different species were elucidated by cryoEM. TRPM2 from the sea anemone (*Nematostella vectensis*) NvTRPM2 lacks the NUDT9H domain, probably due to the flexibility of this part of the molecule.¹⁹ TRPM2 from zebrafish (*Danio rerio*) drTRPM2 has been solved in the apo state and in an ADPR-bound state.²⁰ While ADPR could not be localized in the NUDT9H domain, electron density corresponding to ADPR was found in the *N*-terminal MHR1/2 domain. Comparison of the conformations of the apo and the ADPR-bound state as well as mutational analysis indicate that in drTRPM2 gating of the channel occurs via binding of ADPR to the MHR1/2 domain instead NUDT9H.²⁰ Structures of human TRPM2 (hTRPM2) have been resolved for the apo state, an ADPR bound (primed) state and an open conformation bound to ADPR and Ca²⁺.²¹ Mutagenesis of the MHR1/2 domain and removal of the NUDT9H domain show that in human TRPM2 channel gating occurs after binding of ADPR to the NUDT9H domain, but again poor resolution prevents the placement of ADPR in the presumed binding pocket. Structure-based drug design therefore still awaits a high resolution structure of either the full-length channel or the isolated NUDT9H domain of TRPM2.

Recently, we synthesised ADPR analogues to explore the role of the adenosine, pyrophosphate and terminal ribose motifs in activation of TRPM2.^{22,23} It is of note that the pyrophosphate-forming couplings used generally employed morpholidate- or CDI –mediated methodologies. Yields were generally low and reaction times were generally long and improvements are warranted. To our surprise the majority of structural modifications of ADPR led to compounds that do not activate TRPM2, indicating that all three moieties are required for channel opening.²³ These studies also revealed that hydroxyl group removal at C2' resulted, in the case of ADPR, in a compound that is a TRPM2 agonist with significantly higher efficacy than ADPR itself²³ and also improved the antagonist 8-Ph-ADPR

(IC₅₀ 11 μmol/L compared to 8-Ph-2'-deoxy-ADPR, IC₅₀ 2 μmol/L).²² We also highlighted the importance of the terminal ribose for TRPM2 activation by evaluating a series of modified ADPR analogues.²³ While simple ADP neither activated nor antagonized the channel, introduction of substituents at the β-phosphate that increasingly resembled the terminal ribose (Figure 1) returned antagonist properties, but none of the analogues exhibited agonist activity. The fact that β-(tetrahydrofuran-2-yl)methyl-ADP, an analogue with the ribofuranose backbone, but lacking the hydroxyl groups, did not activate the channel but instead antagonized ADPR indicated that one or more of the hydroxyl groups might be important for the gating of TRPM2. Previous studies have shown hydroxyl group deletion to be a valuable tool in SAR elucidation for complex bioactive molecules.^{24,25,26} While such endeavors may be synthetically protracted in order to effect a desired precision edit to the parent molecule, they can divulge key mechanistic information.^{27,28}

Figure 1. The structure of ADPR and known terminal ribose analogues²⁹

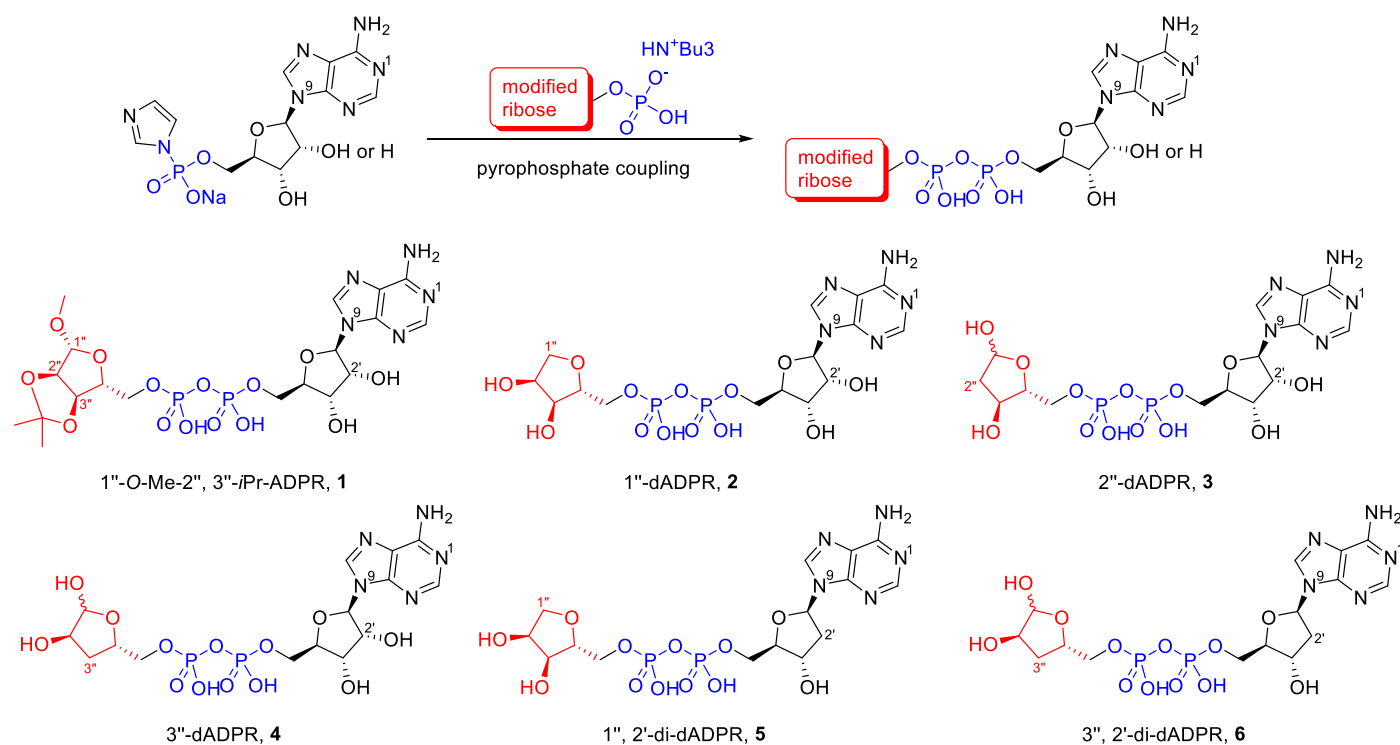


We report here the synthesis and electrophysiological evaluation of a series of novel terminal-ribose-modified analogues of the TRPM2 agonists ADPR and 2'-deoxy-ADPR to study the role of the individual hydroxyl groups of the terminal ribose in TRPM2 activation. The critical step of analogue formation was achieved by combining a sugar phosphate with a P-activated nucleotide to form a single molecule linked by a pyrophosphate bond. A recently reported improved procedure for the preparation of NDP-sugars from nucleoside phosphorimidazolides³⁶ uses 2-4 equivalents of magnesium chloride to achieve high reaction yields and short reactions times for nucleoside 5-phosphorimidazolidine coupling to sugar phosphates.

Results & Discussion

The terminal ribose of ADPR is essential for the ligand-driven activation of the cation channel TRPM2.²⁹ To investigate the SAR of this part of ADPR in more detail, we focused upon the synthesis of all three possible 1''-deoxy, 2''-deoxy and 3''-deoxy terminal ribose ADPR derivatives **2**, **3**, **4** by selective deletion of the appropriate hydroxyl group (Figure 2). The similar reactivity of the ribose hydroxyl groups and potential for ring opening of the cyclic hemi-acetal required development of multi-step synthetic routes with selective masking and deprotection. The modified ribose building blocks were then coupled to either AMP or 2'-deoxy-AMP via pyrophosphate coupling reaction (Figure 2). We also synthesized a corresponding analogue **1** with all hydroxyl groups present, but blocked with small alkyl groups to interrogate the H-bond donating capability of the terminal ribose (Figure 2). The chemically stable analogues were evaluated regarding their agonist and antagonist activity by patch clamp experiments in whole cell configuration using HEK cells with stable expression of human TRPM2.

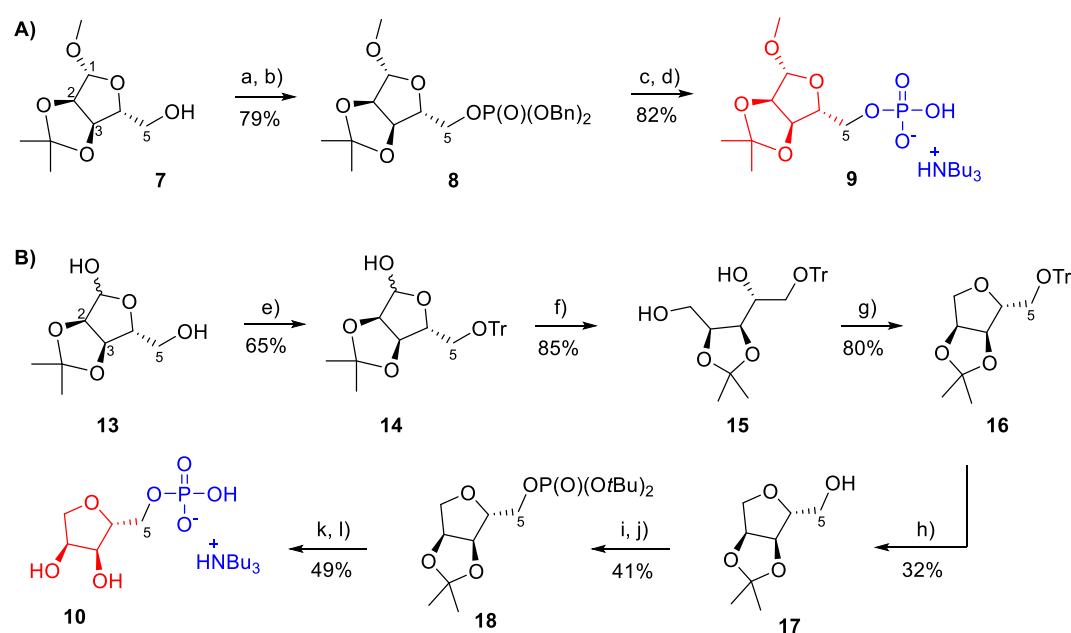
Figure 2. Pyrophosphate coupling reaction and ADPR analogues prepared in this study



At first, routes to the required terminal ribose 5-phosphates were designed. It was envisaged that such phosphates would be ideal coupling partners for activated nucleotide imidazolides. Protected ribose monophosphate **9** was chosen to exploit the ligand space around the terminal ribose, H-bond donating capability, and due to its good susceptibility for coupling with activated nucleosides. The precursor of **9**, compound **8**, was obtained from protected ribose³⁰ **7** by phosphitylation with dibenzyl *N,N*-diisopropylphosphoramidite and subsequent oxidation using hydrogen peroxide³¹ to afford **8** (79%), (Scheme 1 A). Compound **8** was then hydrogenated using Pd/C to afford **9** (82%). Ribose-5-monophosphate **9** was converted to its tributylammonium salts and freeze-dried. Synthesis of 1-deoxyribose-5-phosphate **10**, to explore the role of the anomeric OH group of terminal ribose, started from 2, 3-isopropylidene ribose **13** that was protected at position 5 by a trityl group, to afford **14** (65%) (Scheme 1 B). Reduction of **14** by sodium borohydride led to compound **15** (85%). Reaction of **15**

with tosyl chloride in pyridine gave no significant result at room temperature, but heating the reaction mixture up to 60 °C helped to afford the protected 1-deoxyribose **16** (80%). However, deprotection of **16** with acetic acid failed after multiple attempts, as both the 5-*O*-trityl and 2-3-*O*-isopropylidene protecting groups were cleaved, leading to the undesired fully deprotected 1-deoxyribose or to complex mixtures. Using the alternative deprotection of **16** with formic acid in diethylether³² afforded compound **17** (32%).³³ Phosphitylation and subsequent oxidation of **17** with di-*tert*-butyl *N,N*-diisopropylphosphoramidite gave the phosphate derivative **18** (41%). Careful deprotection of **18** with aqueous trifluoroacetic acid at low temperature afforded the target 1-deoxyribose-5-phosphate **10** (49%) that was subsequently converted to its tributylammonium salt.

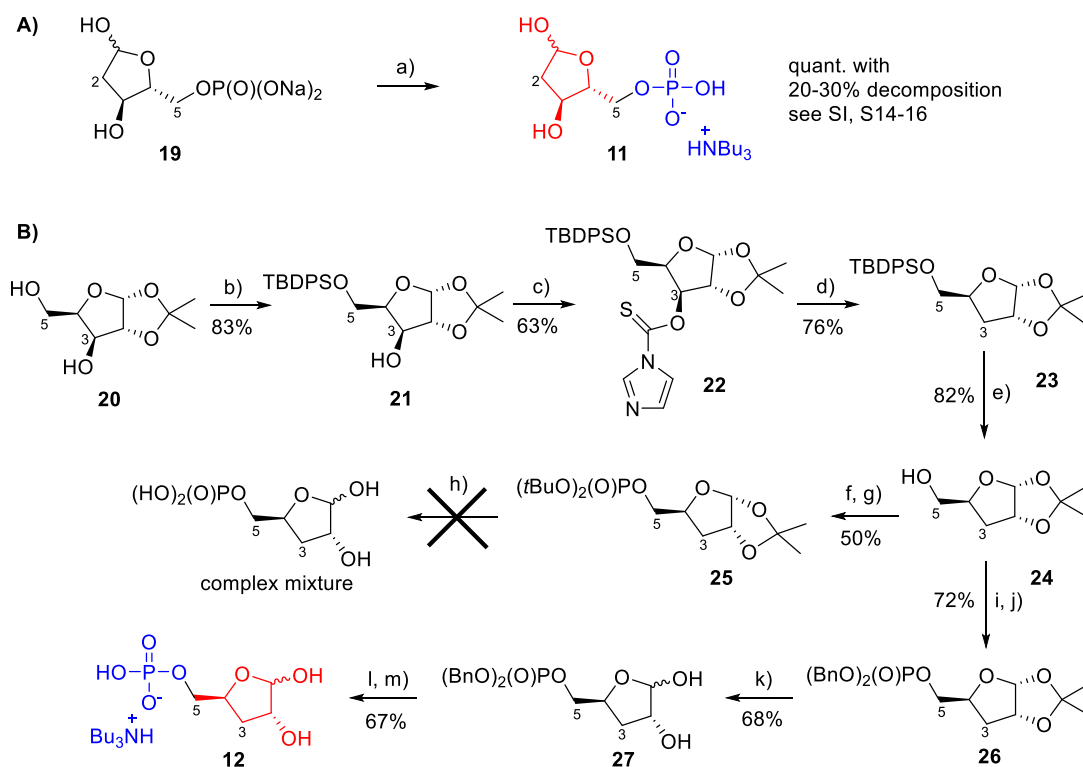
Scheme 1. Synthesis of modified terminal riboses **9** and **10**.



Reagents and conditions: a) 5-Ph-1-*H*-tetrazole, dibenzyl *N,N*-diisopropylphosphoramidite, DCM, 20 °C, 1h; b) triethylamine, H₂O₂, 0-20 °C, 1h; c) hydrogen (balloon), Pd/C, 5h, TEAB (1M), 20 °C; d) Dowex® D50 (H⁺), tributylamine; e) tritylchloride, pyridine, 20 °C, 16h; f) sodium borohydride, ethanol, 0-20 °C, 2h; g) tosylchloride, pyridine, 60 °C, 16h; h) HCOOH/diethylether, 20 °C, 16h; i) 5-Ph-1-*H*-tetrazole, di-*tert*-butyl *N,N*-diisopropylphosphoramidite, DCM, 20 °C, 1h; j) triethylamine, H₂O₂, 0-20 °C, 1h; k) aqueous TFA, 0-20 °C, 4h; l) Dowex® D50 (H⁺), tributylamine.

Synthesis of the 2-deoxy version of the terminal ribose (Scheme 2) started from commercially available 2-deoxyribose-5-phosphate sodium salt **19** (Sigma Aldrich) that was converted to its corresponding mono-tributylammonium salt **11** by using Dowex® resin (H⁺ form) followed by titration with tributylamine to pH~7 (Scheme 2A). Partial decomposition (20-30%) of **19** was observed during the transformation to **11** via ³¹PNMR, suggesting that 2-deoxyribose-5-monophosphate is less stable towards changes in pH (see Supplementary Information, S14-16) compared to its 1-deoxy- and 3-deoxy-counterparts. Synthesis of 3-deoxyribose-5-monophosphate **12** started from 1, 2-*O*-isopropylidene xylose **20**,³⁴ that was protected at the 5-hydroxyl group as the TBDPS ether to give **21** (83%) (Scheme 2B). Reaction of **21** with 1,1'-thiocarbonyldiimidazole in DCM afforded compound **22** (63%). Reductive deoxygenation of **22** with tributyltin hydride led to the compound **23** (76%) which was subsequently deprotected in buffered TBAF to give 1, 2-*O*-isopropylidene-3-deoxyribose **24** (82%). Phosphitylation and subsequent oxidation of **24** gave the desired di-*tert*-butyl phosphate derivative **25** (50%). However, several attempts to deprotect compound **25** with aqueous TFA led to a complex mixture suggesting that di-*tert*-butyl protected phosphate **25** was not the optimal precursor for **12**. Therefore, **24** was phosphitylated and subsequently oxidized to its dibenzylphosphate derivative **26** (72%) that was successfully deprotected to give **27** (68%). Hydrogenation of **27** followed by neutralization of the free phosphate using tributylamine afforded the target analogue 3-deoxyribose-5-phosphate as its mono-tributylammonium salt **12** (67%).

Scheme 2. Synthesis of modified terminal riboses **11** and **12**.

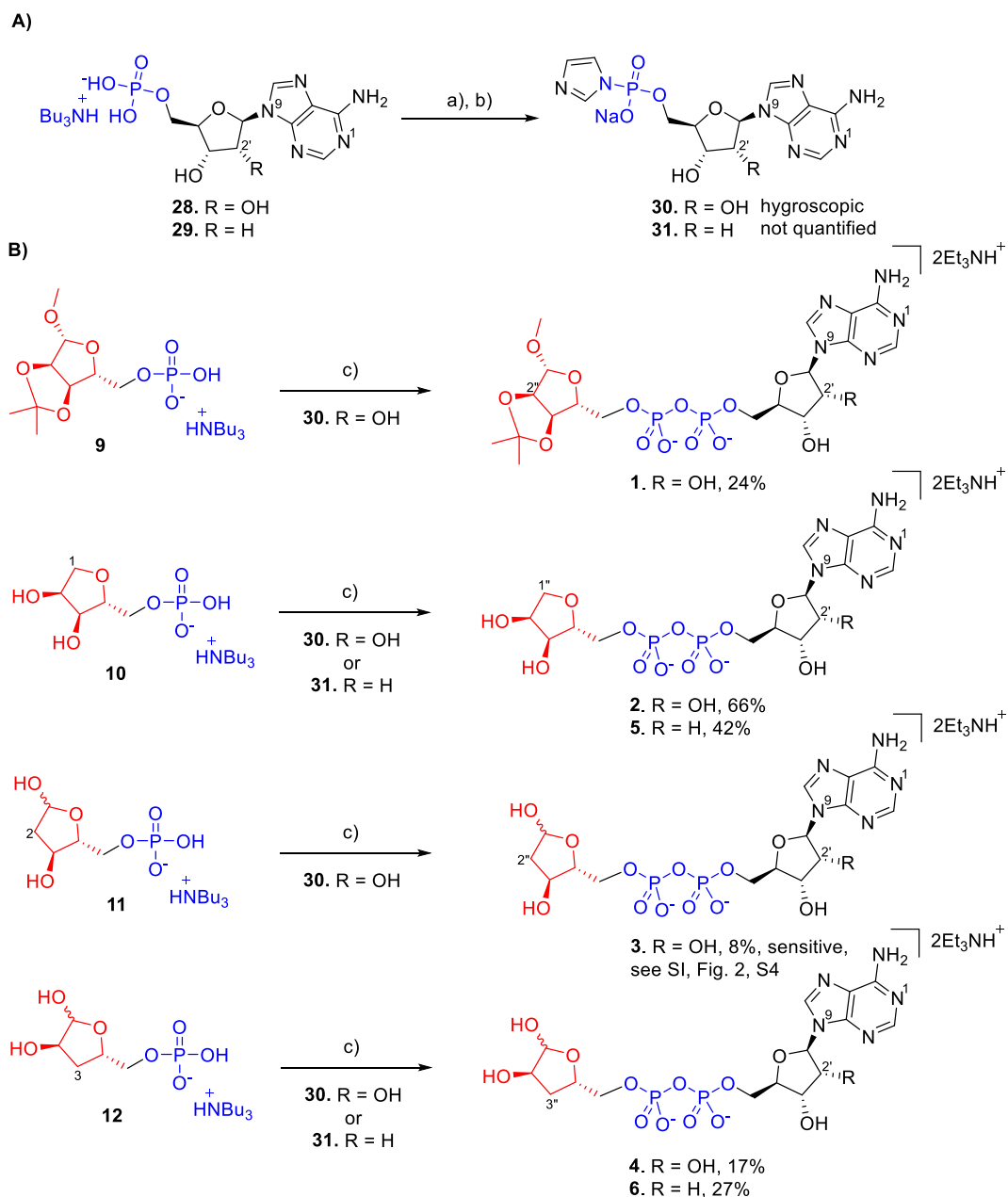


Reagents and conditions: a) Dowex® D50 (H⁺), tributylamine; b) TBDPS chloride, DMAP, Pyridine, 20 °C, 16 h; c) 1,1'-thiocarbonyldiimidazole, DCM, reflux, 2h; d) tributyltinhydride, AIBN, toluene, 116 °C, 3h; e) TBAF.3H₂O, acetic acid, 20 °C, 3h; f) 5-Ph-1-*H*-tetrazole, di-*tert*-butyl *N,N*-diisopropylphosphoramidite, DCM, 20 °C, 1h; g) triethylamine, H₂O₂, 0-20 °C, 1h; h) various conditions with aqueous TFA; i) 5-Ph-1-*H*-tetrazole, dibenzyl *N,N*-diisopropylphosphoramidite, DCM, 20 °C, 1h; j) triethylamine, H₂O₂, 0-20 °C, 1h; k) aqueous TFA, 0 °C, 1.5h; l) hydrogen (balloon), Pd/C, 4h, 20 °C; m) tributylamine.

In previous work we used morpholidate chemistry or 1,1'-carbonyldiimidazole-based methodology in coupling reactions, but these had been less than satisfactory during ADPR analogue preparation. To find the most suitable conditions here for pyrophosphate bond formation between the corresponding ribose-5-monophosphate and either AMP or 2'-dAMP, two different methods were tried. Activation of AMP tributylammonium salt with 1,1'-carbonyldiimidazole³⁵ followed by addition of the triethylammonium salt of ribose **9** did show formation of the desired pyrophosphate product **1** by HPLC. However, several byproducts were also observed (see Supplementary Information Figure 1a,

S2). In contrast, following the Dabrowski-Tumanski procedure³⁶ activation of the AMP tributylammonium salt using imidazole, 2,2'-dithiodipyridine (AldrithiolTM) and triphenyl phosphine cleanly afforded the AMP-imidazolide that was isolated by precipitation with a cold solution of NaI in acetone. When stirred with the triethylammonium salt of **9** and magnesium chloride in DMF, the AMP-imidazolide gave almost exclusively the desired compound **1** (see Supplementary Information Figure 1b, S2-S3). Finally, purified target ribose derivatives **9**, **10**, **11** and **12** were individually coupled to imidazolide-activated AMP or 2'-deoxy-AMP (Scheme 3). Adenosine-5'-monophosphate **28** and 2'-deoxy-adenosine-5'-monophosphate **29** were transformed to their imidazolides **30** and **31**, by reaction of the corresponding mono-tributylammonium salt of **28** and **29** and imidazole using the AldrithiolTM and triphenylphosphine condensation protocol.³⁶ Imidazolides **30** and **31** were precipitated from the reaction mixture by addition of a 0.1 M solution of sodium iodide in cold, anhydrous acetone, and were directly used for coupling. The target modified ADPR analogues **1** - **6** were then prepared by coupling³⁶ the corresponding imidazolide **30** or **31** and mono-tributylammonium salt of the modified ribose-5-phosphate **9**, **10**, **11** or **12** to give variable yields of the desired ADPR analogues **1** (24%), **2** (66%), **3** (8%), **4** (17%), **5** (42%), **6** (27%). Final compounds **1-6** were purified by semi-preparative HPLC using triethylammonium bicarbonate buffer and isolated as the corresponding triethylammonium salts.

Scheme 3. Synthesis of target terminal-modified ADPR analogues



Reagents and conditions: a) AldrithiolTM, imidazole, triethylamine, triphenylphosphine, 20 °C, 16h; b) NaI, acetone (precipitation), 0 °C; c) MgCl₂, dimethylformamide, 20 °C, 3-16h.

Stability of terminal-modified ADPR analogues 1 - 6

The stability of ADPR analogues **1 - 6** was evaluated using analytical HPLC. Aqueous solutions of **1 - 6** stored at 5 °C were compared with freshly prepared standard solutions of **1 - 6** (from solid samples stored at -20 °C). All compounds except **3** were stable in aqueous solutions for several days when

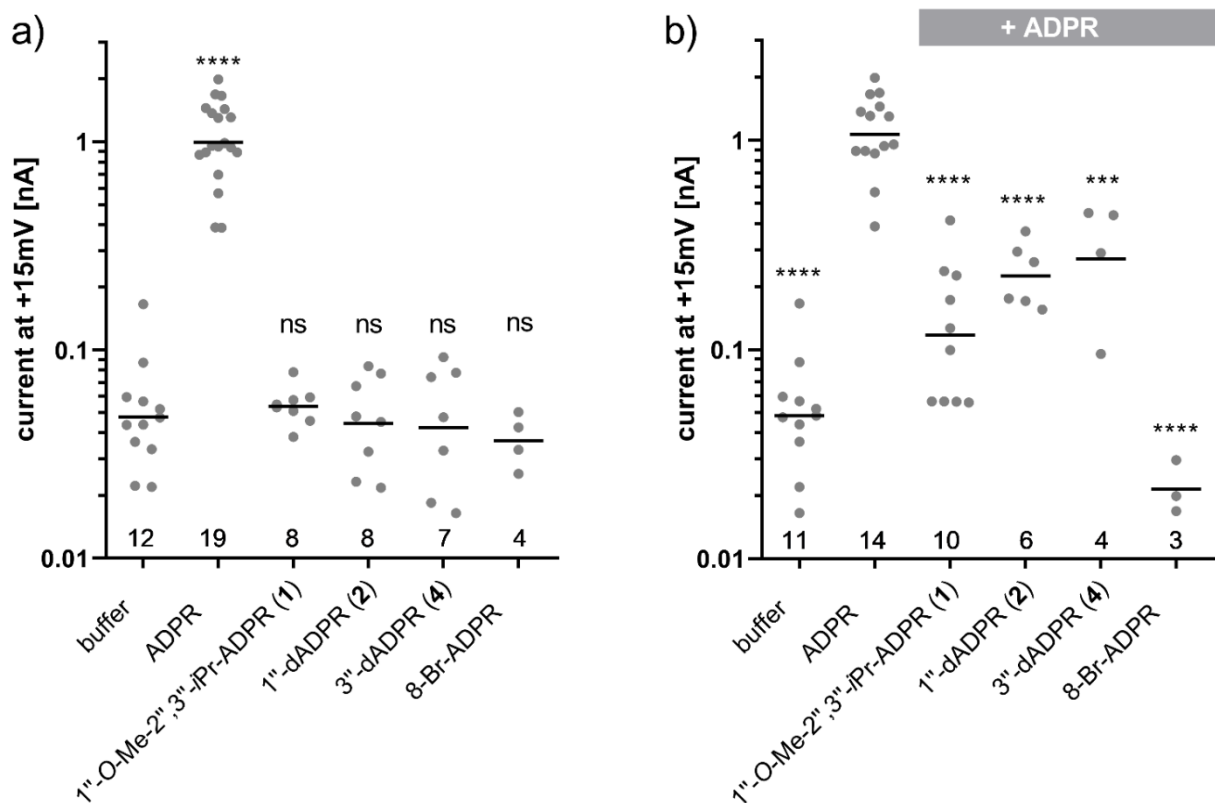
stored in the fridge (5 °C). Compound **3**, 2''-deoxy-ADPR was found to be unstable under these conditions. We also observed decomposition of **3** during transport and sample preparation, so we could not do electrophysiological experiments with **3** (Supplementary Information Figure 2, S4). The instability of **3** may correspond with the lower stability of 2-deoxyribose-5-monophosphate **11** that was sensitive towards changes in pH during conversion of the commercially available sodium salt to the tributylammonium salt (conversion of **19** to **11**). This decomposition led to hydrolysis of the 5'-phosphate (approx. 20-30% de-phosphorylation of **11** was observed via ³¹PNMR). The corresponding decomposition by de-phosphorylation in 2''-deoxy-ADPR **3** might cleave the pyrophosphate bond, and would be predicted to lead to inactive fragments.

Evaluation of novel analogues against TRPM2

Recently, we investigated the importance of the ADPR terminal ribose for the activation of TRPM2.²⁹ This can be investigated by whole cell patch clamp experiments. During these experiments a glass pipette with a diameter much smaller than a cell and filled with a solution mimicking the ionic composition of the cytosol is attached to the plasma membrane. By applying suction to the pipette the membrane patch underneath the pipette is ruptured. In voltage-clamp experiments a voltage is applied between an electrode in the pipette solution and a bath electrode in the bath solution surrounding the cells. This voltage results in ions moving through channels in the membrane resulting in a current that can be recorded.^{37,38} In HEK293 cells expressing TRPM2 addition of ADPR to the pipette solution results in a current that is absent in wild type HEK293 cells. ADPR analogues can be tested for their ability to activate the channel and they can also be tested for antagonist activity by adding them in addition to ADPR. We found that adenosine 5'-monophosphate (AMP) and adenosine 5'-diphosphate (ADP) neither activated or inhibited the channel even when applied in large excess over ADPR.²⁹ Interestingly, replacement of the terminal ribose of ADPR with small substituents led to analogues that did not activate the channel, but inhibited activation by ADPR indicating that they compete with ADPR

for the ligand binding site of TRPM2. Thus, activation of TRPM2 may be attributed to specific interactions between the hydroxyl groups of the terminal ribose of ADPR and the ligand binding NUDT9H domain of TRPM2. We evaluated the deoxy analogues **2** and **4** and an ADPR analogue **1** where all hydroxyl groups of the terminal ribose are masked and could no longer act as hydrogen bond donors (Figure 3). It was not possible to evaluate **3** biologically due to instability issues (see above). During the experiments we included 8-Br-ADPR as a control. 8-Br-ADPR has previously been shown to inhibit activation of TRPM2 by ADPR most likely by competing with the agonist.¹⁰

Figure 3. Effect of ADPR analogues on whole cell currents in TRPM2 expressing HEK293 cells

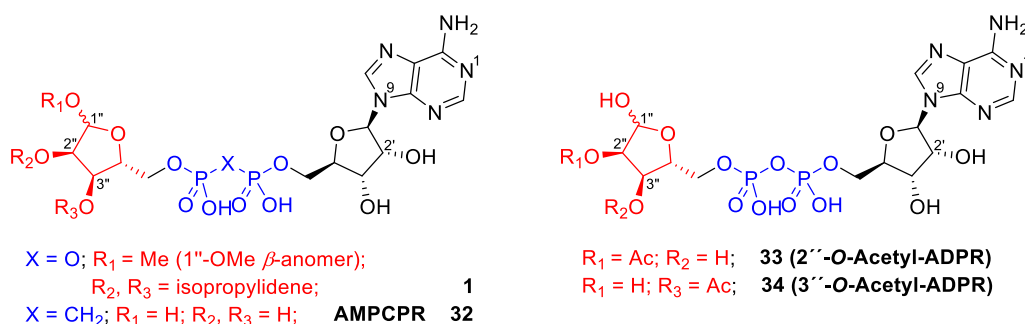


Conditions: a) Effect of ADPR analogues on whole cell currents in TRPM2 expressing HEK293 cells. Experiments were done as outlined in the Experimental Section. ADPR or the indicated ADPR analogue were added to the pipette solution at a concentration of 100 $\mu\text{mol/L}$. b) Effect of ADPR analogues on TRPM2 whole cell currents elicited by ADPR. In this case the pipette solution contained either no nucleotide (buffer), 100 μM ADPR or a combination of 100 μM ADPR with 900 μM of the indicated ADPR analogue. 8-Br-ADPR was included as inhibitor control. Points indicate the maximum

current from individual cells. The number of cells for each condition is indicated at the bottom. Bars represent the mean of the log-transformed currents. (ns = non-significant, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$). Control conditions on both panels include overlapping sets of data since on some days both agonist and antagonist experiments were performed.

As expected, the masked analogue 1''-O-Me-2'', 3''-iPr-ADPR **1** did not activate TRPM2 on its own (Figure 3a). Since it is more space-filling than ADPR we were unsure whether it would compete with ADPR for binding to the NUDT9H domain. When applied in excess over ADPR, however, it indeed inhibited activation of TRPM2 (Figure 3b), showing that further steric constraints than those simply at the 1''-position of the terminal ribose are tolerated and may potentially be exploitable for TRPM2 antagonist optimization.

Figure 4. The structures of masked ADPR **1**, α - β -methylene ADPR (AMPCPR) **32**, 2''-O-acetyl-ADPR **33**, and 3''-O-acetyl-ADPR **34**.



Interestingly, an analogue **32** (Figure 4) of the low-affinity partial TRPM2 agonist α - β -methylene ADPR (AMPCPR)³⁹ masked in the same way as **1** was recently described to be a high affinity TRPM2 antagonist that inhibits ADPR-elicited TRPM2 currents with an IC_{50} of 5.7 μM .⁴⁰ Luo *et al* did not explicitly mention whether they tested this compound for TRPM2 activation but, since it fully inhibits the channel at 100 μM , it is unlikely to exhibit agonist activity. Methylene analogues may show increased stability towards cellular pyrophosphatases like NUDT9 and NUDT5 which may be retained in the cell during whole cell patch-clamp experiments. Despite earlier reports that indicated an ADPR

pyrophosphatase activity of the NUDT9H domain of TRPM2,¹ that might contribute to degradation of agonists with pyrophosphate bridge, this has recently been disputed.³ The observed difference could also result from different experimental conditions as we buffered the intracellular Ca^{2+} concentration to 200 nmol/L with 10 mmol/L of EGTA whereas Luo *et al.* left the Ca^{2+} concentration largely unbuffered (50 $\mu\text{mol/L}$ EGTA) which would result in a much steeper dependence of the current on the agonist concentration and a more pronounced inhibition by the antagonist.

Contrary to our expectations both ADPR analogues **2**, **4** lacking hydroxyl groups at the terminal ribose did not induce significant whole cell currents in TRPM2-expressing HEK cells at 100 $\mu\text{mol/L}$ (Figure 3a). This was unexpected as *O*-acetyl-ADPR, a product of NAD-dependent histone deacetylases of the Sir2 family, is an effective activator of TRPM2 that binds to the NUDTH domain with similar affinity as ADPR, as shown by UV crosslinking experiments and activates TRPM2 with a comparable concentration dependence as ADPR.⁴¹ 2''-*O*-acetyl-ADPR **33**, the product of the deacetylases, undergoes in neutral solution rapid intramolecular transesterification resulting in roughly equal amounts of 2''-*O*-acetyl-ADPR **33** and 3''-*O*-acetyl-ADPR **34**.⁴² Either the nucleotide binding site of TRPM2 is indifferent to the position of the acetyl group, which would indicate that neither the 2''- nor the 3''-hydroxyl group is essential as hydrogen bond donor, or there is a specificity for one of the isomers which would mean that only half of the *O*-acetyl-ADPR molecules in solution are effective agonists. Since the latter seems unlikely, this may suggest that the 2''-OH and 3''-OH groups are hydrogen-bond acceptors, a feature that could be retained even when acetylated. We have previously shown that α -*O*-methyl-ADPR and β -*O*-methyl-ADPR (Figure 1) do not activate TRPM2, but both inhibit channel activation by ADPR.²⁹ This indicates that the 1''-hydroxyl group in ADPR could not be masked or that there is no space in this region to accommodate even a relatively small methyl group, without losing agonist activity in contrast to the 2''- and 3''-hydroxyl groups. So why does 3''-deoxy-ADPR **4** not activate the channel? Possible explanations may be that the conformation of the furanose

ring or the ratio between the two anomeric forms of the ribose are affected by the absence of the 3''-hydroxyl group in a way that prevents the proper hydrogen bonding necessary for channel activation, or that this hydroxyl is required as a hydrogen bond acceptor.

The five-membered ribose sugar ring is puckered due to non-bonded interactions and the energetically most stable conformation has all substituents as far apart as possible.⁴³ Thus, the different substitution patterns in the terminal ribose of ADPR, 1''-deoxy-ADPR **2**, 2''-deoxy-ADPR **3** and 3''-deoxy-ADPR **4**, might be expected to produce differing types of puckering. Extensive analysis of the ribose and 2'-deoxy-ribose rings in RNA and DNA nucleotides has shown that while ribose adopts a primarily C3'-*endo* configuration, 2'-deoxy-ribose adopts a primarily C2'-*endo* form.⁴⁴ Such changes alter the orientation of the ribose ring substituents, and this may affect the orientation of the substituents and the way they are presented to the binding site, as well as the stability of a given ADPR analogue by bringing nucleophilic hydroxyl groups into proximity for intramolecular attack on the pyrophosphate. Unfortunately, the broad nature of the peaks in the ¹H-NMR spectra of the ADPR analogues means that it is not possible fully to determine the coupling constants, and hence to analyze fully each terminal ribose ring conformation and suggests that rapid interconversion between configurations may be taking place in solution.

ADPR is a mixture of terminal ribose α - and β -anomers and it is not known whether one or both anomers activate TRPM2. Analysis of ¹H-NMR spectra demonstrates that both 2''-deoxy-ADPR **3** and 3''-deoxy-ADPR **4** are also mixtures of the α and β -forms in the following ratios (α : β , 2''-deoxy-ADPR **3**, 1:1.17; 3''-deoxy-ADPR **4**; 1:4). Both analogues still adopt cyclic (as opposed to open chain form) structures for the terminal ribose. Given that both anomers (α - and β -forms) are present in both cases, it would seem unlikely that this could be the sole cause of their observed inactivity.

Surprisingly, di-deoxy ADPR analogues **5** and **6** did not have significant antagonist activity (data not shown). This was unexpected, since the potency of our previous antagonist 8-Ph-ADPR ($IC_{50} = 11$ μ M) was improved by the 2'-deoxy modification (8-Ph-2'-deoxy-ADPR $IC_{50} = 3$ μ M).

The recent cryo-EM structures of human TRPM2 show that in the absence of ADPR interactions between the NUDT9H domain and MHR1/2 domains of the same and adjacent subunits lock the channel in a closed conformation.²¹ Binding of ADPR impacts the conformation of the NUDT9H domain of TRPM2 in a way that leads to disengagement of the inter-subunit interactions, priming the channel for full activation by binding of Ca^{2+} ions to a cytosolic site near the pore region²¹. The NUDT9H domain of TRPM2 has a bi-lobed structure with the two lobes forming a cleft that is supposed to bind ADPR. While comparison of the apo and the ADPR bound state shows a narrowing of this cleft upon binding of ADPR²¹, the low local resolution of the structure in the NUDT9H domain does not currently allow for localization of ADPR in this binding pocket or for identification of the interactions, that would trigger the conformational changes leading to the primed state. In the absence of a high resolution structure, SAR data therefore remain essential for ligand based drug design. Our current data further support the role of interactions involving the terminal ribose of ADPR in the gating of TRPM2.

Recently, key differences between hTRPM2 and the invertebrate *Nematostella vectensis* variant NvTRPM2 were identified. Thus, two reported synthetic ADPR analogue hTRPM2 antagonists were NvTRPM2 agonists⁴⁵. Moreover, a regulatory function of NUDT9H in NvTRPM2 opposed to that in hTRPM2 was uncovered through the action of another synthetic analogue, inosine 5'-diphosphate ribose IDPR. Thus, ADPR analogues, such as those reported here, are at the cutting edge of progress to unravel the mechanism of TRPM2 function.

Conclusions

Building on our previous study that showed the essential role of the terminal ribose of ADPR in gating of the Ca^{2+} -permeable, non-selective cation channel TRPM2, we synthesized adenine, ribose and 2'-deoxyribose analogues of ADPR **1-6** each lacking one of the hydroxyl groups of the terminal ribose. The convergent synthesis of the targets **1-6** consisted of two separate steps: a) synthesis of particular deoxyribose monophosphates **9-12** and b) coupling analogues **9-12** to the AMP and 2'-deoxy-AMP imidazolides **30-31**, following the Dabrowski-Tumanski procedure³⁶. Such an approach seems to be highly efficient in preparation of such complex molecules as modified ADPR analogues, offering improved yields in our hands compared to pyrophosphate formation using AMP-morpholides or activation using CDI. While 2''-deoxy-ADPR **3** proved to be unstable, analogues **1, 2, 4, 5, 6** showed acceptable stability and were tested electrophysiologically in patch-clamp cell experiments. Neither 1''-deoxy-ADPR **2** nor 3''-deoxy-ADPR **4** was able to activate TRPM2 significantly; instead, both **2** and **4** were weak antagonists of ADPR-mediated TRPM2 activation in whole cell experiments, further highlighting the sensitivity of channel activation to structural changes of the terminal ribose. 1''-O-Me-2'', 3''-iPr-ADPR, 1''-deoxy-, and 3''-deoxy-ADPR (**1, 2** and **4**) were all antagonists of ADPR-mediated Ca^{2+} -release in whole cell patch clamp experiments. Unlike in our previous observations, where including an additional 2'-deoxy-modification generated a more potent TRPM2 antagonist, the corresponding 1''- and 3'', 2'-dideoxy-analogues **5, 6** were less potent antagonists of TRPM2. Results further highlight the significance of the ADPR terminal ribose for activation of TRPM2 and the use of synthetic ADPR analogues in general. The synthesis of such ADPR analogues as chemical biology tools using the methods outlined here is proving invaluable in the wider TRPM2 field.⁴⁵ Further analogues to interrogate individual hydroxyl group stereochemistry or individually mask each of the three terminal hydroxyl groups to probe hydrogen bonding interactions at the TRPM2 binding site will undoubtedly shed more light on the role of the terminal ribose.

Experimental Section

General

All reagents and solvents were of commercial quality and were used without further purification, unless described otherwise. Triethylamine was dried over potassium hydroxide, distilled and then stored over potassium hydroxide pellets. H₂O was of MilliQ quality. Unless otherwise stated, all reactions were carried out under an inert atmosphere of nitrogen. All ¹H, ¹³C, and ³¹P NMR spectra of the final compounds were collected, either on a Varian Mercury 400 MHz or Bruker Avance III 500 MHz Spectrometer. Chemical shifts (δ) are reported in parts per million (ppm) and all ¹H- and ¹³C-NMR assignments are based on COSY, HSQC, HMBC, and DEPT experiments. Abbreviations for splitting patterns are as follows: br, broad; s, singlet; d, doublet; t, triplet; m, multiplet etc. High resolution time-of-flight mass spectra were obtained on a Bruker Daltonics micrOTOF mass spectrometer using electrospray ionisation (ESI). Analytical HPLC analyses were carried out on a Waters 2695 Alliance module equipped with a Waters 2996 Photodiode Array Detector (210 – 350 nm). The chromatographic system consisted of a Hichrom Guard Column for HPLC and a Phenomenex Synergi 4u MAX-RP 80A column (150 × 4.60 mm), eluted at 1 mL/min with a gradient of MeCN in 0.05 M TEAB. Semi-preparative HPLC was carried out on a Waters 2525 pump with manual FlexInject. The chromatographic system consisted of a Phenomenex Gemini, 5u, C18, 110A column (250 x 10.00 mm), eluted at 5 mL/min.

1-*O*-Methyl-2, 3-*O*-isopropylidene-D-ribofuranose (7).³⁰ Compound **7** was prepared according to the literature from D-ribose. Yield (4.28 g, 63%). NMR and HRMS confirmed. ¹HNMR (400MHz, CDCl₃) δ 4.93 (s, 1H, H-1), 4.78 (d, *J* = 6.0 Hz, 1H, H-2), 4.54 (d, *J* = 6.0 Hz, 1H, H-3), 4.38 (bs, 1H, H-4), 3.66-3.55 (m, 2H, H-5_{a, b}), 3.38 (s, 3H, OCH₃), 3.19 (bs, 1H, OH), 1.44 (s, 3H, CH₃), 1.27 (s, 3H, CH₃).

Dibenzyl 1-*O*-methyl-2,3-*O*-isopropylidene- β -D-ribofuranose-5-phosphate (8). 1-*O*-methyl-2,3-*O*-isopropylidene- β -D-ribofuranose **7** (100 mg, 0.49 mmol) and 5-phenyl-1*H*-tetrazole (143 mg, 0.98 mmol) were co-evaporated with dry toluene (2 \times). Then the solid mixture was suspended in dichloromethane (2 mL) and dibenzyl *N,N*-diisopropylphosphoramidite (250 μ L, 0.669 mmol) was added dropwise. The mixture was stirred at 20 °C for 2 hours. TLC analysis (petroleum ether/EtOAc, 2:1 v/v) showed complete phosphitylation. The reaction was cooled to 0 °C and triethylamine (407 μ L, 2.92 mmol) was added followed by aqueous hydrogen peroxide (35%, 109 μ L, 1.24 mmol). The resulting mixture was stirred at 20 °C for 1 h. The mixture was diluted with EtOAc (20 mL) and extracted with aqueous Na₂SO₃ (10%, w/v). The organic phase was dried (Na₂SO₄), solids were removed by filtration and the solvent was evaporated in *vacuo*. The crude product was purified by Isco-Flash chromatography using petroleum ether/EtOAc (1:0 \rightarrow 0:1, v/v). This procedure afforded the title compound **8** as a colorless liquid (180 mg, 79%). ¹HNMR (400MHz, CDCl₃) δ 7.40-7.31 (m, 10H, Bn-2, 3, 4), 5.10-5.0 (m, 4H, Bn-CH₂), 4.93 (s, 1H, H-1), 4.60 (dd, 1H, *J* = 6.0 Hz, *J* = 0.4 Hz, H-2), 4.50 (d, 1H, *J* = 6.0 Hz, H-3), 4.28 (dt, 1H, *J* = 6.0 Hz, *J* = 0.4 Hz, H-4), 4.0-3.88 (m, 2H, H-5), 3.26 (s, 3H, OMe), 1.46 and 1.28 (2 \times s, 2 \times 3H, CH₃*i*Pr). ³¹PNMR (160MHz, CDCl₃) δ -1.23 (s, P). ¹³CNMR (100MHz, CDCl₃) δ 135.7 and 135.6 (2 \times C-*ipso*-Bn), 128.5 and 127.9 (10 \times C-Bn-*o, m, p*), 112.5 (C(CH₃)), 109.3 (C-1), 84.9 (C-3), 84.6 (d, *J* = 9.0 Hz, C-4), 81.4 (C-2), 69.4 and 69.4 (CH₂-Bn), 67.1 (d, *J* = 6.0 Hz, C-5), 55.0 (OMe), 26.3 and 24.9 (CH₃*i*Pr). HRMS (ES⁺) calcd for C₂₃H₂₉NaO₈P 487.1492 [M+Na]⁺, found 487.1501.

1-*O*-Methyl-2,3-*O*-isopropylidene- β -D-ribofuranose-5-phosphate (9). Dibenzyl 1-*O*-methyl-2,3-*O*-isopropylidene- β -D-ribofuranose-5-phosphate **8** (180 mg, 0.39 mmol) was dissolved in MeOH-water (17:3 v/v, 8 mL). After vacuum/argon deoxygenation of the reaction mixture the catalytic amount of palladium on charcoal (10%, 20 mg) was added. The reaction was stirred under positive pressure of hydrogen atmosphere (balloon) at 20 °C for 5 hours. Then the reaction was flushed with argon and neutralized by addition of triethylammonium bicarbonate buffer (TEAB, 1 mL, 1M). The

solvent was evaporated and the crude product was co-evaporated (2 ×) with dry toluene and dried under high-vacuum to give a colourless solid film, compound **9** (1.17 × Et₃N salt, 122 mg, 82%). ¹H NMR (400 MHz, CD₃OD) δ 4.83 (s, 1H, H-1), 4.77 (m, 1H, H-2), 4.53 (d, 1H, *J* = 8.0 Hz, H-3), 4.24-4.18 (m, 1H, H-4), 3.83-3.72 (m, 2H, H-5), 3.24 (s, 3H, OMe), 3.08 (q, 7H, *J* = 8.0 Hz, NCH₂CH₃), 1.37 (t, 3H, CH₃*i*Pr), 1.27-1.21 (m, 13.5 H, NCH₂CH₃, CH₃*i*Pr). ³¹P NMR (160 MHz, *d*₄-MeOD) δ 0.87 (s, P). ¹³C NMR (100 MHz, CD₃OD) δ 113.3 (C(CH₃)), 110.7 (C-1), 86.9 (d, *J* = 9.0 Hz, C-4), 86.4 (C-3), 83.4 (C-2), 66.3 (d, *J* = 5.0 Hz, C-5), 55.1 (OMe), 47.5 (NCH₂CH₃), 26.7 and 25.0 (CH₃*i*Pr), 9.2 (NCH₂CH₃). HRMS (ES⁻) calcd for C₉H₁₆O₈P 283.0588 [M-H]⁻, found 283.0575. (*The product was transformed into its tributylammonium salt before the pyrophosphate coupling.*)

Transformation of 9 to mono-tributylammonium salt: The triethylammonium salt of compound **9** (89 mg, 0.23 mmol) was dissolved in water (2 mL) and treated with Dowex® D50 (H⁺). Then the resin was removed by filtration and the solution of **9** (free acid) was titrated with tributylamine (54 μL, 0.23 mmol, 1 equivalent, pH 7). The solution of **9** was freeze-dried overnight to obtain a colorless film, compound **9** (mono-tributylammonium salt, quant.). ¹H NMR (400 MHz, H₂O, water suppression) δ 5.09 (s, 1H, H-1), 4.90 (d, 1H, *J* = 5.6 Hz, H-2), 4.65 (d, 1H, *suppressed by H₂O*, H-3), 4.43-4.35 (m, 1H, H-4), 3.89-3.82 (m, 2H, H-5), 3.37 (s, 1H, OMe), 3.15-3.07 (m, 6H, NCH₂), 1.69-1.59 (m, 6H, NCH₂CH₂), 1.49 (s, 3H, CH₃*i*Pr), 1.40-1.30 (m, 9H, N(CH₂)₂CH₃, CH₃*i*Pr), 0.95-0.87 (m, 9H, N(CH₂)₃CH₃). ³¹P NMR (160 MHz, D₂O) δ 0.14 (s, P). ¹³C {¹H} NMR (100 MHz, D₂O) δ 113.2 (C(CH₃)), 108.5 (C-1), 85.0 (d, *J* = 9.0 Hz, C-4, HSQC), 86.4 (C-3), 81.0 (C-2), 65.2 (d, *J* = 7.0 Hz, C-5, HSQC), 54.8 (OMe), 52.8 (NCH₂), 25.3 (CH₃*i*Pr), 25.3 (NCH₂CH₂), 23.7 (CH₃*i*Pr), 19.3 (N(CH₂)₂CH₂), 12.8 (N(CH₂)₃CH₃).

2, 3-*O*-Isopropylidene-5-*O*-trityl-D-ribofuranose (14**).** 2,3-*O*-Isopropylideneribose **13** (4 g, 21.1 mmol) was dissolved in pyridine (10 mL). Trityl chloride (7.05 g, 25.3 mmol) was added to the stirred solution and the mixture was stirred at 20 °C for 16 hours. Water (30 mL) was added and the mixture was stirred additional 10 min. The aqueous phase was extracted with dichloromethane (3×). The

combined organic phase was dried with MgSO_4 , solids were removed by filtration, and the solvent was evaporated in *vacuo*. The crude product was purified by silica gel chromatography petroleum ether/EtOAc (1:0 \rightarrow 0:1, v/v). This procedure afforded the title compound **14** as a white amorphous solid (5.9 g, 65%). ^1H NMR²⁵ (400MHz, d_6 -DMSO) δ 7.42-7.32 and 7.30-7.24 (2 \times m, 12 + 3H, *o*, *m*, *p*-Tr), 6.41 (d, 1H, J = 4.0 Hz, OH), 5.15 (d, 1H, J = 4.0 Hz, H-1), 4.56 (dd, 1H, J = 6.0 Hz, J = 1.0 Hz, H-2), 4.36 (d, 1H, J = 6.0 Hz, H-3), 4.11-4.05 (m, 1H, H-4), 3.16-3.05 (m, 2H, H-5), 1.37 and 1.23 (2 \times s, 2 \times 3H, CH_3iPr). HRMS (ES^+) calcd for $\text{C}_{27}\text{H}_{28}\text{NaO}_5$ 455.1829 $[\text{M}+\text{Na}]^+$, found 455.1842.

2, 3-*O*-Isopropylidene-5-*O*-trityl-D-ribitol (15). 2, 3-*O*-Isopropylidene-5-*O*-trityl-D-ribofuranose **14** (5.2 g, 11.98 mmol) was dissolved in EtOH and cooled to 0 °C. Sodium borohydride (460 mg, 12.1 mmol) was added to the solution in three portions over the period of 30 min. The mixture was stirred at 20 °C for 2 h. Water (80 mL) was added and the solution was carefully acidified with acetic acid to pH 6. Then the aqueous phase was extracted with dichloromethane (3 \times). The combined organic phase was dried (Na_2SO_4), solids were removed by filtration and the solvent was evaporated in *vacuo*. The crude product was purified by Isco-Flash chromatography using CH_2Cl_2 /Acetone (1:0 \rightarrow 1:1, v/v). This procedure afforded the title compound **15** as an amorphous white solid (4.35 g, 83%). ^1H NMR (400MHz, d_6 -DMSO) δ 7.48-7.41 (m, 6H, *o*-Tr), 7.35-7.28 (m, 6H, *m*-Tr), 7.26-7.20 (m, 3H, *p*-Tr), 5.15 (d, J = 5.6 Hz, OH-4), 4.80 (t, J = 5.6 Hz, OH-1), 4.17-4.10 (m, 1H, H-2), 4.08-4.01 (m, 1H, H-3), 3.78-3.66 (m, 2H, H-4 and H-1_a), 3.51-3.43 (m, 1H, H-1_b), 3.15-3.11 (m, 1H, H-5_a), 3.06-3.01 (m, 1H, H-5_b), 1.20 (s, 6H, CH_3iPr). $^{13}\text{C}\{^1\text{H}\}$ NMR (100MHz, d_6 -DMSO) δ 144.0 (3 \times C-*ipso*-Tr), 128.4 (6 \times C-*o*-Tr), 127.7 (6 \times C-*m*-Tr), 126.8 (3 \times C-*p*-Tr), 107.4 (C(CH_3)), 85.7 (C(Tr)), 78.2 (C-2), 76.4 (C-3), 68.2 (C-4), 66.1 (C-1), 60.0 (C-5), 27.8 and 25.4 (CH_3iPr). HRMS (ES^+) calcd for $\text{C}_{27}\text{H}_{30}\text{NaO}_5$ 457.1985 $[\text{M}+\text{Na}]^+$, found 457.2004.

1-Deoxy-2, 3-*O*-isopropylidene-5-*O*-trityl-D-ribofuranose (16). 2,3-*O*-Isopropylidene-5-*O*-trityl-D-ribitol **15** (1.89 g, 4.35 mmol) was dissolved in pyridine (15 mL) and tosyl chloride (2.49 g, 13.06 mmol) was added. The mixture was stirred at 60 °C for 16 h. TLC analysis showed the reaction was

complete. The tosyl chloride surplus was decomposed by addition of water (50 mL) and the aqueous phase was extracted with dichloromethane (3×). The combined organic phase was dried (Na₂SO₄), solids were removed by filtration and the solvent was evaporated in *vacuo*. The crude product was purified by Isco-Flash chromatography using petroleum ether/EtOAc (1:0 → 1:1, v/v), affording the title compound **16** as an amorphous white solid (1.44 g, 80%). ¹HNMR (400MHz, CDCl₃) δ 7.44-7.39 (m, 6H, *o*-Tr), 7.33-7.27 (m, 6H, *m*-Tr), 7.26-7.21 (m, 3H, *p*-Tr), 4.91-4.86 (m, 1H, H-2), 4.65 (dd, 1H, *J* = 6.4 Hz, *J* = 1.2 Hz, H-3), 4.24-4.19 (m, 1H, H-4), 4.15 (dd, 1H, *J* = 10.4 Hz, *J* = 4.4 Hz, H-1_a), 4.05 (dd, 1H, *J* = 10.0 Hz, *J* = 1.0 Hz, H-1_b), 3.27 (dd, 1H, *J* = 10.0 Hz, *J* = 4.0 Hz, H-5_a), 3.11 (dd, 1H, *J* = 10.0 Hz, *J* = 4.4 Hz, H-5_b), 1.51 and 1.34 (2 × s, 6H, CH₃*i*Pr). ¹³C{¹H}NMR (100MHz, CDCl₃) δ 143.6 (3 × C-*ipso*-Tr), 128.6 (6 × C-*o*-Tr), 127.8 (6 × C-*m*-Tr), 127.2 (3 × C-*p*-Tr), 112.3 (C(CH₃)), 87.1 (C(Tr)), 84.0 (C-4), 83.0 (C-3), 81.6 (C-2), 74.3 (C-1), 64.5 (C-5), 26.6 and 25.1 (CH₃*i*Pr). HRMS (ES⁺) calcd for C₂₇H₂₈NaO₄ 439.1880 [M+Na]⁺, found 439.1868.

1-Deoxy-2,3-*O*-isopropylidene-D-ribofuranose (17). 1-Deoxy-2,3-*O*-isopropylidene-5-*O*-trityl-D-ribofuranose **16** (100 mg, 0.24 mmol) was dissolved in diethylether-HCOOH (2 mL, 1:1, v/v) and stirred at 20 °C for 16 h. TLC analysis (petroleum ether/EtOAc, 1:1, v/v) indicated the complete conversion of reaction and the mixture was evaporated to dryness. The dry residue was suspended in water and the suspension was filtered through a cotton pad. Evaporation of water afforded the title compound **17** as a colorless viscous liquid (14.5 mg, 32%). ¹HNMR (400MHz, CDCl₃) δ 4.83-4.78 (m, 1H, H-2), 4.59 (dd, 1H, *J* = 6.4 Hz, *J* = 2.0 Hz, H-3), 4.13-4.09 (m, 1H, H-4), 3.98-3.95 (m, 2H, H-1_{a, b}), 3.65 (dd, 1H, *J* = 11.6 Hz, *J* = 4.0 Hz, H-5_a), 3.58 (dd, 1H, *J* = 11.6 Hz, *J* = 6.4 Hz, H-5_b), 1.51 and 1.33 (2 × s, 6H, CH₃*i*Pr). ¹³C{¹H}NMR (100MHz, CDCl₃) δ 113.0 (C(CH₃)), 84.8 (C-4), 81.8 (C-3), 81.0 (C-2), 72.8 (C-1), 61.8 (C-5), 26.7 and 25.0 (CH₃*i*Pr). HRMS (ES⁺) calcd for C₈H₁₄NaO₄ 197.0784 [M+Na]⁺, found 197.0791.

1-Deoxy-2,3-*O*-isopropylidene-D-ribofuranose-5-*O*-di-*tert*-butylphosphate (18). 1-deoxy-2,3-*O*-isopropylidene-D-ribofuranose **17** (94 mg, 0.49 mmol) and 5-phenyl-1*H*-tetrazole (145 mg, 0.99 mmol) were co-evaporated with dry toluene (2×). Then, the dry mixture was dissolved in dichloromethane (2 mL) and di-*tert*-butyl *N,N*-diisopropylphosphoramidite (234 μL, 0.74 mmol) was added dropwise. The mixture was stirred at 20 °C for 2 hours after which TLC analysis (petroleum ether/EtOAc, 2:1, v/v) showed the reaction was complete. It was cooled to 0 °C and triethylamine (411 μL, 2.97 mmol) added, followed by hydrogen peroxide (35% aq., 109 μL, 1.24 mmol). The resulting mixture was stirred at 20 °C for 1 hour. The mixture was diluted with EtOAc (20 mL) and extracted with aqueous Na₂SO₃ (10%, w/v). The organic phase was dried (Na₂SO₄), solids were filtered off and the solvent was evaporated in *vacuo*. The crude product was purified by Isco-Flash chromatography using petroleum ether/EtOAc (1:0 → 0:1, v/v), to afford the title compound **18** as a colorless liquid (74 mg, 41%). ¹H NMR (400 MHz, CDCl₃) δ 4.85-4.81 (m, 1H, H-2), 4.76 (dd, 1H, *J* = 6.4 Hz, *J* = 1.2 Hz, H-3), 4.23-4.18 (m, 1H, H-4), 4.2-3.94 (m, 4H, H-1, H-5), 1.51 (s, 3H, CH₃*i*Pr), 1.48 (s, 18H, CH₃*t*Bu), 1.33 (s, 3H, CH₃*i*Pr). ³¹P NMR (160 MHz, CDCl₃) δ -10.1 (s, P). ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 112.6 (C(CH₃)), 83.1 (d, *J* = 8.0 Hz, C(*t*Bu)), 82.6 (d, *J* = 7.0 Hz, C-4), 82.3 (C-3), 81.2 (C-2), 73.9 (C-1), 66.6 (d, *J* = 7.0 Hz, C-5), 29.8-29.7 (m, CH₃*t*Bu), 26.5 and 24.9 (CH₃*i*Pr). HRMS (ES⁺) calcd for C₁₆H₃₁NaO₇P 389.1700 [M+Na]⁺, found 389.1699.

Tributylammonium 1-deoxy-2,3-*O*-isopropylidene-D-ribofuranose-5-phosphate (10). Di-*tert*-butyl 1-deoxy-2,3-*O*-isopropylidene-D-ribofuranose-5-phosphate **18** (36 mg, 0.098 mmol) was dissolved in aqueous methanol (1:1, v/v, 2 mL) and the solution was cooled to 0 °C. Trifluoroacetic acid (2 mL) was added dropwise and the solution was allowed to warm up to 20 °C and stirred for an additional 4 h. The solution was evaporated to dryness and the residue co-evaporated with water (3×) and with methanol (3×). The procedure afforded the pure phosphate derivative as a colorless gum (21 mg, quantitative) which was directly dissolved in water (2 mL), neutralized with tributylamine (33 μL, 0.14 mmol), the solvent evaporated and the residue co-evaporated with EtOH (2×). The title compound

10 was obtained as the mono-tributylammonium salt (18.7 mg, 49%). ^1H NMR (400MHz, MeOH) δ 4.21-4.16 (m, 1H, H-2), 4.12-4.07 (m, 1H, H-3), 4.03-3.87 (m, 4H, H-1_a, 4, 5_{a,b}), 3.69 (dd, 1H, $J = 9.6$ Hz, $J = 4.0$ Hz, H-1_b), 3.10-3.03 (m, 6H, NCH₂), 1.73-1.64 (m, 6H, NCH₂CH₂), 1.41 (m, 6H, N(CH₂)₂CH₂), 1.00 (t, 9H, $J = 7.6$ Hz, CH₃Bu). ^{31}P NMR (160MHz, MeOH) δ 1.09 (s, P). $^{13}\text{C}\{^1\text{H}\}$ NMR (100MHz, MeOH) δ 83.3 (d, $J = 8.0$ Hz, C-4), 73.7 (C-1), 73.6 (C-3), 72.5 (C-2), 66.3 (d, $J = 5.0$ Hz, C-5), 54.0 (NCH₂), 27.0 (NCH₂CH₂), 21.0 (N(CH₂)₂CH₂), 14.0 (CH₃Bu). HRMS (ES⁻) calcd for C₅H₁₁O₇P 213.0170 [M-H]⁻, found 213.0162.

2-Deoxy-D-ribofuranose-5-phosphate sodium salt (19). Compound **19** (sodium salt) was purchased from Sigma Aldrich (UK) and found to be a mixture of α and β anomers in the ratio 1/1.17 α/β . ^1H NMR (500MHz, D₂O) δ 5.52-5.47 (m, 1.6H, H-1 α , β , deuterium exchange), 4.51-4.46 (m, 1H, H-3 β), 4.38-4.34 (m, 1H, H-3 α), 4.25-4.20 (m, 1H, 4 α), 4.04-3.99 (m, 1H, H-4 β), 3.85-3.81 (m, 2H, H-5 β), 3.81-3.75 (m, 2H, H-5 α), 2.51-2.42 (m, 1H, H-2 α), 2.22-2.14 (m, 2H, H-2 β), 1.91-1.83 (m, 1H, H-2 α). ^{31}P NMR (202MHz, D₂O) δ 3.65 (P- β) and 3.68 (P- α). $^{13}\text{C}\{^1\text{H}\}$ NMR (126MHz, D₂O) δ 98.4 (C-1 β), 98.0 (C-1 α), 84.9 (d, $J = 10.08$ Hz, C-4 β), 84.6 (d, $J = 8.82$ Hz, C-4 α), 71.6 (C-3 β), 71.1 (C-3 α), 64.6 (d, $J = 5.0$ Hz, C-5 β), 63.7 (d, $J = 3.8$ Hz, C-5 α), 40.8 (C-2 β), 40.6 (C-2 α). Compound **19** was transformed to its tributylammonium salt.

2-Deoxyribose-5-phosphate mono-tributylammonium salt (11). Commercially available 2-deoxyribose monophosphate sodium salt **19** (Sigma Aldrich) (8 mg, 0.029 mmol) was dissolved in water (2 mL) and acidified to pH 1-2 by using the ion-exchange resin Dowex® (D50 H⁺). The resin was filtered off and the free phosphate was neutralized by addition of one equivalent of tributylamine (6.9 μL , 0.029 mmol). The aqueous solution was freeze-dried (quant.). (*NMR analysis showed partial decomposition (approx. 20-30%), of 2'-deoxyribose). The compound itself is sensitive to basic conditions (pH 9).*) ^1H NMR (500MHz, D₂O) δ 5.50-5.47 (m, 0.54H, H-1 β), 5.43-5.41 (dd, 0.46H, $J = 5.5$ Hz, $J = 2.5$ Hz, H-1 α), 4.43-4.39 (m, 0.53H, H-3 β), 4.29-4.25 (m, 0.47H, H-3 α), 4.15-4.1 (m, 0.46H, H-4 α), 4.02-3.97 (m, 0.7H, H-5 β), 3.94-3.85 (m, 1.32H, H-4 β , 5 α , 5 β), 3.84-3.77 (m, 0.52H,

H-5 α), 3.09-3.01 (m, 10H, NCH₂(CH₂)₂CH₃), 2.38-2.30 (m, 0.5H, H-2 α), 2.12-1.99 (m, 1H, H-2 β), 1.86-1.79 (m, 0.5H, H-2 α), 1.72-1.60 (m, 10H, NCH₂CH₂CH₂CH₃), 1.47-1.35 (m, 10H, NCH₂CH₂CH₂CH₃), 1.04-0.94 (m, 15H, NCH₂(CH₂)₂CH₃). ³¹P NMR (202MHz, D₂O) δ 1.10 (P- β) and 1.03 (P- α). ¹³C {¹H} NMR (126MHz, D₂O) δ 100.0 (C-1 β), 99.5 (C-1 α), 86.3 (d, J = 8.4 Hz, C-4 β), 85.6 (d, J = 8.7 Hz, C-4 α), 73.5 (C-3 β), 73.0 (C-3 α), 67.3 (d, J = 5.4 Hz, C-5 β), 66.1 (d, J = 5.3 Hz, C-5 α), 53.8 (NCH₂(CH₂)₂CH₃), 43.0 (C-2 β), 42.6 (C-2 α), 26.9 (NCH₂CH₂CH₂CH₃), 21.1 (NCH₂CH₂CH₂CH₃), 14.0 (NCH₂(CH₂)₂CH₃).

1, 2-*O*-Isopropylidene-D-xylofuranose (20).³⁴ Finely powdered D-xylose (10 g, 67 mmol) was suspended in acetone (260 mL) containing sulphuric acid (10 mL) and stirred for 30 min until dissolved. The solution was cooled to 0 °C and a solution of Na₂CO₃ (13 g in 112 mL H₂O) was added. The mixture was stirred at 20 °C for 1 hour and then solid Na₂CO₃ (7 g) was added. The slurry was stirred at 20 °C for an further 30 min. Solids were removed by filtration and washed with acetone. Acetone was evaporated and the water phase was extracted with EtOAc (3 \times) and dried with Na₂SO₄. Isco-Flash chromatography using petroleum ether/EtOAc (1:0 \rightarrow 1:1, v/v) afforded the title compound **20** as a colorless viscous oil (4.35 g, 34%) and doubly protected 1, 2-*O*-isopropylidene-3,5-*O*-isopropylidene-D-xylofuranose (3.29 g, 21%).³⁴ ¹H NMR (400MHz, CDCl₃) δ 5.98 (d, 1H, J = 3.6 Hz, H-1), 4.52 (d, 1H, J = 3.6 Hz, H-2), 4.32 (d, 1H, J = 2.4 Hz, H-4), 4.19-4.11 (m, 2H, H-3, 5_a), 4.05 (dd, 1H, J = 12.4 Hz, J = 2.4 Hz, H-5_b), 1.48 and 1.32 (CH₃*i*Pr). ¹³C {¹H} NMR (100MHz, CDCl₃) δ 112.0 (C(CH₃)₂), 105.1 (C-1), 85.9 (C-2), 78.7 (C-3), 77.3 (C-4), 61.5 (C-5), 26.9 and 26.3 (CH₃*i*Pr). HRMS (ES⁺) calcd for C₈H₁₄NaO₅ 213.0733 [M+Na]⁺, found 213.0787.

5-*O*-*tert*-Butyl(diphenyl)silyl-1, 2-*O*-isopropylidene-D-xylofuranose (21). 1, 2-*O*-Isopropylidene-D-xylofuranose **20** (3.63 g, 19.08 mmol) and DMAP (5 mg) were dissolved in pyridine (15 mL). *tert*-Butyl(diphenyl)silyl chloride (4.89 mL, 19.08 mmol) was added dropwise and the solution was stirred at 20 °C for 16 hours. Solvent was evaporated and the crude product was purified by Isco-Flash chromatography using Petroleum ether/EtOAc (1:0 \rightarrow 1:1, v/v). This procedure afforded the title

compound **21** as a white solid (6.58 g, 83%). ^1H NMR (400MHz, CDCl_3) δ 7.74-7.65 (m, 4H, *o*-Ph), 7.48-7.38 (m, 6H, *m*, *p*-Ph), 6.00 (d, 1H, $J = 3.6$ Hz, H-1), 4.54 (d, 1H, $J = 3.6$ Hz, H-2), 4.39-4.35 (m, 1H, H-4), 4.16-4.12 (m, 1H, H-3), 4.12-4.09 (m, 2H, H-5), 3.99 (d, 1H, $J = 2.4$ Hz, 3-OH), 1.47 and 1.33 (CH_3iPr), 1.06 (CH_3tBu). $^{13}\text{C}\{^1\text{H}\}$ NMR (100MHz, CDCl_3) δ 135.8 and 135.7 (*o*-Ph), 132.6 and 132.1 (*C-ipso*-Ph), 130.2 and 128.1 ($2 \times \text{d}$, $J = 2.2$ Hz, $J = 1.7$ Hz, *m*, *p*-Ph), 111.8 ($\text{C}(\text{CH}_3)_2$), 105.2 (*C*-1), 85.6 (*C*-2), 78.6 (*C*-3), 77.0 (*C*-4, obstructed by CDCl_3 signal), 62.9 (*C*-5), 27.0 and 26.3 (CH_3iPr), 26.9 (CH_3tBu), 19.2 ($\text{C}(\text{CH}_3)_3$). HRMS (ES^+) calcd for $\text{C}_{24}\text{H}_{32}\text{NaO}_5\text{Si}$ 451.1911 $[\text{M}+\text{H}]^+$, found 451.1929.

5-*O*-tert-Butyl(diphenyl)silyl-1, 2-*O*-isopropylidene-3-thiocarbonylimidazolo-D-xylofuranose (22). A mixture of 5-*O*-tert-butyl(diphenyl)silyl-1, 2-*O*-isopropylidene-D-xylofuranose **21** (3.0 g, 7.01 mmol) and 1, 1'-thiocarbonyldiimidazole (2.37 g, 13.32 mmol) was gently refluxed in dichloroethane at 85 °C for 2 hours. The solvent was evaporated and the crude product was purified by Isco-Flash chromatography using petroleum ether/EtOAc (1:0 \rightarrow 0:1, v/v) to afford the title compound **22** as a white solid (2.4 g, 63%). ^1H NMR (400MHz, CDCl_3) δ 8.24-8.21 (m, 1H, Imidazole), 7.64-7.60 (m, 2H, Ph), 7.55-7.51 (m, 2H, Ph), 7.47 (t, 1H, $J = 1.6$ Hz, Imidazole), 7.46-7.33 (m, 4H, Ph), 7.29-7.23 (m, 2H, Ph), 7.04 (q, 1H, $J = 0.8$ Hz, Imidazole), 5.98 (d, 1H, $J = 2.8$ Hz, H-3), 5.94 (d, 1H, $J = 4.0$ Hz, H-1), 4.72 (d, 1H, $J = 4.0$ Hz, H-2), 4.61 (dq, 1H, $J = 8.8$ Hz, $J = 2.8$ Hz, H-4), 4.02 (dd, 1H, $J = 10.0$ Hz, $J = 4.4$ Hz, H-5_a), 3.82 (dd, 1H, $J = 10.0$ Hz, $J = 8.4$ Hz, H-5_b), 1.60 and 1.36 ($2 \times \text{s}$, $2 \times 3\text{H}$, CH_3iPr), 1.02 (s, 9H, CH_3tBu). $^{13}\text{C}\{^1\text{H}\}$ NMR (100MHz, CDCl_3) δ 182.6 ($\text{C}=\text{S}$), 136.9 (Imidazole), 135.6 and 135.4 (Ph), 132.8 and 132.7 (*C-ipso*-Ph), 131.2 (Imidazole), 130.1 (Ph), 127.9 and 127.8 (Ph), 117.8 (Imz), 112.8 ($\text{C}(\text{CH}_3)_2$), 105.0 (*C*-1), 84.2 (*C*-3), 82.7 (*C*-2), 78.9 (*C*-4), 60.3 (*C*-5), 26.9 (CH_3tBu), 26.7 and 26.4 (CH_3iPr), 19.2 ($\text{C}(\text{CH}_3)_3$). HRMS (ES^+) calcd for $\text{C}_{28}\text{H}_{34}\text{N}_2\text{NaO}_5\text{SSi}$ 561.1850 $[\text{M}+\text{Na}]^+$, found 561.1837.

5-*O*-tert-Butyl(diphenyl)silyl-1, 2-*O*-isopropylidene-3-deoxy-D-ribofuranose (23). Tributyltin hydride (1.07 mL, 3.99 mmol) was added into boiling toluene (90 mL) under argon followed by AIBN

(13 mg, 0.08 mmol). Then 5-*O*-*tert*-butyl(diphenyl)silyl-1, 2-*O*-isopropylidene-3-thiocarbonylimidazolo-D-xylofuranose **22** (1.26 g, 2.35 mmol) was added dropwise in toluene (20 mL). The mixture was refluxed for 3 hours. Solvent was evaporated and the crude product was purified by Isco-Flash chromatography using petroleum ether/EtOAc (1:0 → 0:1, v/v) to afford the title compound **23** as a white solid (731 mg, 76%). ¹HNMR (400MHz, CDCl₃) δ 7.71 (m, 4H, *o*-Ph), 7.45-7.35 (m, 6H, *m*, *p*-Ph), 5.83 (d, 1H, *J* = 3.6 Hz, H-1), 4.75 (t, 1H, *J* = 4.0 Hz, H-2), 4.33 (dq, 1H, *J* = 10.4 Hz, *J* = 4.0 Hz, H-4), 3.82 (dd, 1H, *J* = 11.2 Hz, *J* = 4.4 Hz, H-5_a), 3.76 (dd, 1H, *J* = 11.2 Hz, *J* = 4.0 Hz, H-5_b), 3.08 (dd, 1H, *J* = 13.6 Hz, *J* = 4.8 Hz, H-3_a), 1.88 (ddd, 1H, *J* = 13.6 Hz, *J* = 10.4 Hz, *J* = 4.8 Hz, H-3_b), 1.52 and 1.34 (2 × s, 2 × 3H, CH₃*i*Pr), 1.06 (s, 9H, CH₃*t*Bu). ¹³C{¹H}NMR (100MHz, CDCl₃) δ 135.8 and 135.7 (*o*-Ph), 133.7 and 133.5 (*C*-*ipso*-Ph), 129.8 and 129.8 (*p*-Ph), 127.8 and 127.8 (*m*-Ph), 111.3 (C(CH₃)₂), 105.9 (C-1), 80.6 (C-2), 78.7 (C-4), 64.9 (C-5), 35.1 (C-3), 27.1 (CH₃*i*Pr), 27.0 (CH₃*t*Bu), 26.5 (CH₃*i*Pr), 19.4 (C(CH₃)₃). HRMS (ES⁺) calcd for C₂₄H₃₂NaO₄Si 435.1962 [M+Na]⁺, found 435.1995.

1, 2-*O*-Isopropylidene-3-deoxy-D-ribofuranose (24). A mixture of TBAF·3H₂O (1.55 g, 5.0 mmol) and acetic acid (300 μL, 5.25 mmol) in DMF (10 mL) was stirred at room temperature for 30 min. The mixture was cooled to 0 °C and 5-*O*-*tert*-butyl(diphenyl)silyl-1, 2-*O*-isopropylidene-3-deoxy-D-ribofuranose **23** (687 mg, 1.67 mmol) was added dropwise in DMF (5 mL). The resulting mixture was stirred at 20 °C for 3 hours. Solvent was evaporated and the crude product was purified by Isco-Flash chromatography using petroleum ether/EtOAc (1:0 → 0:1, v/v). This procedure afforded the title compound **24** as white solid (260 mg, 82%). ¹HNMR (400MHz, CDCl₃) δ 5.79 (d, 1H, *J* = 3.6Hz, H-1_α), 4.73 (t, 1H, *J* = 4.4 Hz, H-2), 4.31 (dtd, 1H, *J* = 10.8 Hz, *J* = 4.4 Hz, *J* = 3.2 Hz, H-4), 3.84 (dd, 1H, *J* = 12.0 Hz, *J* = 2.8 Hz, H-5_a), 3.54 (dd, 1H, *J* = 12.0 Hz, *J* = 4.4 Hz, H-5_b), 1.97 (dd, 1H, *J* = 13.6 Hz, *J* = 4.8 Hz, H-3_a), 1.85-1.76 (m, 1H, H-3_b), 1.48 and 1.29 (2 × s, 2 × 3H, CH₃*i*Pr). ¹³C{¹H}NMR (100MHz, CDCl₃) δ 111.4 (C(CH₃)₂), 105.7 (C-1_α), 80.9 (C-2), 78.7 (C-4), 63.1 (C-5), 34.0 (C-3), 26.9 and 26.3 (CH₃*i*Pr). HRMS (ES⁺) calcd for C₈H₁₄NaO₄ 197.0784 [M+Na]⁺, found 197.0793.

1, 2-*O*-Isopropylidene-3-deoxy-D-ribofuranose-5-*O*-di-*tert*-butylphosphate (25). 1, 2-*O*-Isopropylidene-3-deoxy-D-ribofuranose **24** (100 mg, 0.52 mmol) and 5-phenyl-1*H*-tetrazole (154 mg, 1.05 mmol) were co-evaporated with dry toluene (2×). The solid mixture was then dissolved in dichloromethane (2 mL) and di-*tert*-butyl *N, N*-diisopropylphosphoramidite (249 μL, 0.79 mmol) was added dropwise. The mixture was stirred at 20 °C for 2 h. TLC analysis (petroleum ether/EtOAc, 2:1, v/v) showed complete phosphitylation. The reaction was cooled to 0 °C and triethylamine (437 μL, 3.15 mmol) added followed by hydrogen peroxide (35% aq., 116 μL, 1.32 mmol). The resulting mixture was stirred at 20 °C for 1 hour. The mixture was diluted with EtOAc (20 mL) and extracted with aqueous Na₂SO₃ (10%, w/v). The organic phase was dried (Na₂SO₄), solids were filtered off and the solvent was evaporated in *vacuo*. The crude product was purified by Isco-Flash chromatography using petroleum ether/EtOAc (1:0 → 0:1, v/v), followed by second chromatography in dichloromethane/acetone (1:0 → 1:1, v/v). This procedure afforded the title compound **25** as a colourless viscous liquid (97 mg, 50%). ¹HNMR (400MHz, CDCl₃) δ 5.81 (d, 1H, *J* = 4.0 Hz, H-1), 4.73 (t, 1H, *J* = 4.4 Hz, H-2), 4.04 (ddd, 1H, *J* = 8.8 Hz, *J* = 4.4 Hz, *J* = 1.2 Hz, H-4), 4.12-4.06 (m, 1H, H-5a), 4.02-3.96 (m, 1H, H-5b), 2.1 (dd, 1H, *J* = 13.6 Hz, *J* = 4.8 Hz, H-3a), 1.84-1.76 (m, 1H, H-3b), 1.50 (s, 3H, CH₃*i*Pr), 1.48 (s, 9H, CH₃*t*Bu) 1.31 (s, 3H, CH₃*i*Pr). ³¹PNMR (160MHz, CDCl₃) δ -9.91 (s, phosphate). ¹³C{¹H}NMR (100MHz, CDCl₃) δ 111.4 (C(CH₃)₂), 105.8 (C-1), 82.7 and 82.6 (2 × d, *J* = 2.1 Hz, C(CH₃)₃), 80.7 (C-2), 76.6 (d, *J* = 8.7 Hz, C-4), 67.0 (d, *J* = 6.0 Hz, C-5), 35.2 (C-3), 30.01 and 29.97 (2 × CH₃*t*Bu), 27.0 and 26.4 (2 × CH₃*i*Pr). HRMS (ES⁺) calcd for C₁₆H₃₂O₇P 367.1880 [M+H]⁺, found 367.1877; calcd for C₁₆H₃₁NaO₇P 389.1700 [M+Na]⁺, found 389.1737.

1, 2-*O*-Isopropylidene-3-deoxy-D-ribofuranose-5-*O*-dibenzylphosphate (26). 1, 2-*O*-Isopropylidene-3-deoxy-D-ribofuranose **24** (120 mg, 0.63 mmol) and 5-phenyl-1*H*-tetrazole (185 mg, 1.26 mmol) were co-evaporated with dry toluene (2×). Then the solid mixture was dissolved in dichloromethane (3 mL) and dibenzyl *N, N*-diisopropylphosphoramidite (353 μL, 0.95 mmol) was added dropwise. The mixture was stirred at 20 °C for 2 h. TLC analysis (petroleum ether/EtOAc, 2/1,

v/v) showed complete phosphitylation. The reaction was cooled to 0 °C and triethylamine (524 μ L, 3.79 mmol) added followed by hydrogen peroxide (35% aq., 139 μ L, 1.58 mmol). The resulting mixture was stirred at 20 °C for 1 h. The mixture was diluted with EtOAc (20 mL) and extracted with Na₂SO₃ (10% aq., w/v). The organic phase was dried (Na₂SO₄), solids were removed by filtration and the solvent was evaporated in *vacuo*. The crude product was purified by Isco-Flash chromatography using dichloromethane/acetone (1:0 \rightarrow 1:1, v/v) followed by a second chromatography in petroleum ether/EtOAc (1:0 \rightarrow 0:1, v/v) to afford the title compound **26** as a colorless viscous liquid (218 mg, 73%). ¹HNMR (400MHz, CDCl₃) δ 7.37-7.32 (m, 10H, Bn), 5.76 (d, J = 3.6 Hz, H-1), 5.10-5.00 (m, 4H, CH₂Bn), 4.68 (t, 1H, J = 4.4 Hz, H-2), 4.35 (ddd, 1H, J = 8.8 Hz, J = 4.4 Hz, J = 0.8 Hz, H-4), 4.16 (dq, 1H, J = 3.6 Hz, J = 2.8 Hz, H-5_a), 4.00 (dq, 1H, J = 4.4 Hz, J = 2.8 Hz, H-5_b), 2.00 (dd, 1H, J = 13.2 Hz, J = 4.4 Hz, H-3_a), 1.72-1.64 (m, 1H, H-3_b), 1.48 and 1.31 (2 \times s, 2 \times 3H, CH₃*i*Pr). ³¹PNMR (160MHz, CDCl₃) δ -1.00 (s, phosphate). ¹³C{¹H}NMR (100MHz, CDCl₃) δ 136.0 (d, J = 2.0 Hz, C-*ipso*-Bn), 135.9 (d, J = 3.0 Hz, C-*ipso*-Bn), 128.8-128.7 and 128.2-128.1 (2 \times m, C-Bn-*o*, *m*, *p*), 111.5 (C(CH₃)₂), 105.8 (C-1), 80.5 (C-2), 76.4 (d, J = 8.0 Hz, C-4), 69.6 and 69.5 (2 \times d, J = 1.3 Hz and J = 1.4 Hz, CH₂Bn), 68.8 (d, J = 5.6 Hz, C-5), 34.7 (C-3), 27.0 and 26.3 (CH₃*i*Pr). HRMS (ES⁺) calcd for C₂₂H₂₈O₇P 435.1567 [M+H]⁺, found 435.1571; calcd for C₂₂H₂₇NaO₇P 457.1387 [M+Na]⁺, found 457.1392.

3-Deoxy-D-ribofuranose-5-O-dibenzylphosphate (27). An aqueous solution (4 mL) of 1, 2-*O*-isopropylidene-3-deoxy-D-ribofuranose-5-*O*-dibenzylphosphate **26** (67mg, 0.154 mmol) was cooled to 0 °C and trifluoroacetic acid (4 mL) was added. The solution was stirred at 0 °C for 1.5 h and then allowed to warm to 20 °C. The solvent was evaporated in *vacuo* at 30 °C and co-evaporated with water. Crude material was purified by silica gel chromatography using petroleum ether/EtOAc (1:0 \rightarrow 0:1, v/v) to give a colorless solid (mixture of α/β anomers 1:4, 41 mg, 68%). ¹HNMR (400MHz, MeOH) δ 7.39-7.35 (m, 10H, Bn), 5.18 (s, 0.2H, C-1 α), 5.17 (s, 0.8H, C-1 β), 5.09-5.04 (m, 4H, CH₂Bn), 4.45-4.39 (m, 0.8H, H-4 β), 4.39-4.35 (m, 0.2H, H-4 α), 4.13-4.07 (m, 1H, H-2 α , β), 4.06-3.99 (m, 1.8H, H-

5 α_a , β_a , b), 3.90 (ddd, 0.2H, $J = 12.8$ Hz, $J = 5.6$ Hz, $J = 4.0$ Hz, H-5 α_b), 1.96-1.85 (m, 2H, H-3 α , β). ^{31}P NMR (160MHz, MeOH) δ -1.30 (s, α -phosphate), -1.54 (s, β -phosphate). $^{13}\text{C}\{^1\text{H}\}$ NMR (100MHz, MeOH) δ 137.2 and 137.2 (2 x C-*ipso*-Bn), 129.8-129.7 and 129.3-129.2 (2 x m, 10 x C-Bn-*o*, *m*, *p*), 104.3 (C-1 β), 98.4 (C-1 α), 78.6 (d, $J = 6.2$ Hz, C-4 β), 77.1 (C-2 β), 75.8 (d, $J = 6.2$ Hz, C-4 α), 72.4 (d, $J = 4.8$ Hz, C-5 β), 72.2 (C-2 α), 71.0 (d, $J = 4.8$ Hz, C-5 α), 70.9-70.8 (m, **CH**₂Bn), 34.7 (C-3 β), 33.9 (C-3 α). HRMS (ES⁺) calcd for C₁₉H₂₄O₇P 395.1254 [M+H]⁺, found 395.1256; calcd for C₁₉H₂₃NaO₇P 417.1074 [M+Na]⁺, found 417.1079.

3-Deoxy-D-ribofuranose-5-phosphate tributylammonium salt (12). 3-Deoxy-D-ribofuranose-5-*O*-dibenzylphosphate **27** (39 mg, 0.098 mmol) was dissolved in MeOH-water (8 mL, 17:3 v/v). After vacuum/argon deoxygenation of the reaction mixture, palladium on charcoal (Pd/C 10%, 10 mg) was added. The reaction was stirred under positive pressure of hydrogen atmosphere (balloon) at 20 °C for 4 h. Then the reaction was flushed with argon and solids were filtered off and washed with water. The aqueous solution (pH 1-2) was carefully titrated by portionwise addition of tributylamine (5 x 4 μ L, 0.084 mmol, pH 6-7). The solvent was evaporated and the crude product was co-evaporated with dry toluene (2x) and dried on the high-vac pump for 16 h to give a colorless solid film, compound **12** (mixture of α/β anomers 1:4, 1 x Bu₃N salt, 29 mg, 67%). ^1H NMR (500MHz, MeOD) δ 5.20 (d, 0.2H, $J = 4.5$ Hz, H-1 α), 5.12 (s, 0.8H, H-1 β), 4.48-4.41 (m, 0.8H, H-4 β), 4.41-4.36 (m, 0.2H, H-4 α), 4.25-4.20 (m, 0.2H, H-2 α), 4.08 (d, 0.8H, $J = 4.5$ Hz, H-2 β), 3.96-3.84 (m, 1.6H, H-5 β), 3.83-3.79 (m, 0.4H, H-5 α), 3.14-3.08 (m, 6H, NCH₂), 2.12 (ddd, 0.2H, $J = 12.5$ Hz, $J = 5.5$ Hz, $J = 2.5$ Hz, H-3 α_a), 2.04 (ddd, 0.8H, $J = 13.5$ Hz, $J = 8.5$ Hz, $J = 5.0$ Hz, H-3 β_a), 2.01-1.97 (m, 0.2H, H-3 α_b), 0.92 (dd, 0.8H, $J = 14.0$ Hz, $J = 7.0$ Hz, H-3 β_a), 1.73-1.64 (m, 6H, NCH₂CH₂), 1.47-1.38 (m, 6H, N(CH₂)₂CH₂), 10.14 (t, 9H, $J = 7.5$ Hz, N(CH₂)₃CH₃). ^{31}P NMR (202MHz, MeOD) δ 1.04 (s, α -phosphate), 0.97 (s, β -phosphate). $^{13}\text{C}\{^1\text{H}\}$ NMR (126MHz, MeOD) δ 104.3 (C-1 β), 98.2 (C-1 α), 79.7 (d, $J = 8.8$ Hz, C-4 β), 77.4 (C-2 β), 76.8 (d, $J = 8.8$ Hz, C-4 α), 72.3 (C-2 α), 69.7 (d, $J = 5.03$ Hz, C-5 β), 68.4 (d, $J = 5.0$ Hz,

C-5 α), 53.8 (NCH₂), 35.3 (C-3 β), 34.3 (C-3 α), 26.8 (NCH₂CH₂), 21.0 (N(CH₂)₂CH₂), 14.0 (N(CH₂)₃CH₃). HRMS (ES⁻) calcd for C₅H₁₀O₇P 213.0170 [M-H]⁻, found 213.0159.

AMP imidazolidine: Adenosine-5'-phosphoryl imidazolidine (30) AMP sodium salt (50 mg, 0.128 mmol, Sigma Aldrich) was dissolved in water (2 mL) and treated with Dowex® D50 (H⁺) until all the phosphate was transformed to free acid (pH 1-2). Then the resin was filtered off, washed with water, and the solution of **AMP** (free acid) was titrated with tributylamine (30 μ L, 0.13 mmol, 1 equivalent, pH 7). The solution of adenosine-5'-phosphate mono-tributylammonium salt (**28**) was freeze-dried to obtain a light white powder (quantitative).

AMP tributylammonium salt (15 mg, 0.028 mmol) and imidazole (19 mg, 0.28 mmol) were co-evaporated with EtOH (2 \times) and with toluene (2 \times). AldrithiolTM (18.6 mg, 0.085 mmol) was added followed by DMF (220 μ L) and the suspension stirred until all solid matter was dissolved. Triethylamine (16 μ L, 0.113 mmol, distilled-dry-stored over KOH in the dark) and triphenylphosphine (22.2 mg, 0.085 mmol) were added and the solution was stirred at 20 °C for 16 h. **Product precipitation:** The reaction mixture was cooled to 5 °C and a cold solution of NaI (33.8 mg) in dry acetone (2.2 mL) was added at 5 °C. The white precipitate was filtered off and washed with cold acetone. The product was dried under high vacuum for 16 h and stored under argon (crude compound, hygroscopic, yield was not quantified). ¹HNMR (400MHz, *d*₄-MeOD) δ 8.38 (s, H-8), 8.19 (s, H-2), 7.88-7.87 (m, 1H, Imidazole), 7.28-7.26 (m, 1H, Imidazole), 6.99-6.97 (m, 1H, Imidazole), 6.04 (d, 1H, *J* = 6.0 Hz, H-1'), 4.65 (t, 1H, *J* = 5.6 Hz, H-2'), 4.25 (dd, 1H, *J* = 5.2 Hz, *J* = 3.6 Hz, H-3'), 4.18-4.13 (m, 1H, H-4'), 4.06-3.96 (m, 2H, H-5'_{a, b}). ³¹PNMR (160MHz, *d*₄-MeOH) δ -9.03 (s, P). ¹³C{¹H}NMR (100MHz, *d*₄-MeOD) δ 157.4 (C-6), 153.9 (C-2), 150.9 (C-4), 140.9 (C-8), 140.8 (d, *J* = 5.5 Hz, Imidazole), 129.7 (d, *J* = 10.4 Hz, Imidazole), 121.3 (d, *J* = 5.7 Hz, Imidazole), 120.3 (C-5), 89.2 (C-1'), 85.3 (d, *J* = 8.8 Hz, C-4'), 75.8 (C-2'), 72.19 (C-3'), 66.8 (d, *J* = 5.9 Hz, C-5'). HRMS (ES⁻) calcd for C₁₃H₁₅N₇O₆P 396.0827 [M-H]⁻, found 396.0853.

1''- α -O-Methyl-2'',3''-O-isopropylidene adenosine diphosphoriboside (1). Compound **9** (4.3 mg, 0.009 mmol) and MgCl_2 (1.8 mg, 0.018 mmol) were dissolved in DMF (1 mL), co-evaporated with dry toluene (2 \times) and evaporated to dryness. Dry DMF (500 μL) was added to the solid residue and the mixture was stirred at 20 $^\circ\text{C}$ until all solids were dissolved (5-10 min). Then imidazolide **30** (3.5 mg, 0.008 mmol) was added dropwise to the mixture in DMF (400 μL). The mixture was stirred at 20 $^\circ\text{C}$ for 3 h, after which conversion to product was 80-90% by HPLC). Solvent was evaporated in *vacuo* and crude product was dissolved in triethylammonium bicarbonate buffer (TEAB, 3 mL, 0.1M) and purified by semi-preparative, reverse-phase HPLC using a gradient of TEAB (0.1 M)-acetonitrile (95:5 \rightarrow 35:65, v/v). The title compound **1** was obtained as a colorless glass, $1.7 \times \text{Et}_3\text{N}$ salt, (2 μmol , 1.57 mg, 24%). ^1H NMR (500MHz, D_2O) δ 8.48 (s, 1H, H-8), 8.20 (s, 1H, H-2), 6.06 (d, 1H, $J = 5.0$ Hz, H-1'), 4.89 (s, 1H, H-1''), 4.70-4.63 (m, 2H, H-2', 2''), 4.50 (d, 1H, $J = 5.0$ Hz, H-3''), 4.45 (t, 1H, $J = 5.0$ Hz, H-3'), 4.33-4.28 (m, 1H, H-4'), 4.21-4.11 (m, 3H, H-4'', 5'_{a,b}), 3.82-3.75 (m, 2H, H-5''_{a,b}), 3.21 (s, 3H, OMe), 3.12 (m, 9H, NCH_2CH_3), 1.31 (s, 3H, CH_3 -*i*Pr), 1.19 (m, 16.5H, NCH_2CH_3 , CH_3 -*i*Pr). ^{31}P NMR (202MHz, D_2O) δ -11.47 (m, pyrophosphate). $^{13}\text{C}\{^1\text{H}\}$ NMR (126MHz, D_2O) δ 155.1 (C-6), 152.0 (C-2), 149.1 (C-4), 140.0 (C-8), 118.3 (C-5), 112.9 ($\text{C}(\text{CH}_3)_2$), 108.4 (C-1''), 87.0 (C-1'), 84.7 (d, $J = 10.1$ Hz, C-4''), 84.1 (C-3''), 83.9 (d, $J = 6.3$ Hz, C-4'), 80.9 (C-2''), 74.3 (C-2'), 70.3 (C-3'), 65.8 (d, $J = 3.8$ Hz, C-5''), 65.1 (d, $J = 6.3$ Hz, C-5'), 54.7 (OMe), 46.6 (NCH_2CH_3), 25.1 and 23.5 (CH_3 -*i*Pr), 8.2 (NCH_2CH_3). HRMS (ES^-) calcd for $\text{C}_{19}\text{H}_{28}\text{N}_5\text{O}_{14}\text{P}_2$ 612.1114 $[\text{M}-\text{H}]^-$, found 612.1143. UV (H_2O , pH 7.4) λ_{max} 259 nm (ϵ 16900 L/mol.cm). HPLC $t_{\text{R}} = 5.52$ min.

1''-Deoxyadenosine diphosphoriboside (1''-deoxy-ADPR) (2). 1-Deoxy-D-ribofuranose-5-phosphate tributylammonium salt **10** (5.8 mg, 0.0145 mmol) and MgCl_2 (2.8 mg, 0.029 mmol) were dissolved in DMF (1 mL), co-evaporated with dry toluene (2 \times) and evaporated to dryness. Dry DMF (500 μL) was added to the solid residue and the mixture was stirred at 20 $^\circ\text{C}$ until all solids were dissolved (5-10 min). Then AMP-imidazolide **30** (5.5 mg, 0.013 mmol) was added dropwise to the mixture in DMF (400 μL). The mixture was stirred at 20 $^\circ\text{C}$ for 3 h. Solvent was evaporated in *vacuo*

and the crude product was dissolved in triethylammonium bicarbonate buffer (TEAB, 3 mL, 0.1M) and purified by semi-preparative, reverse-phase HPLC using a gradient of TEAB (0.1M)-acetonitrile (95:5 \rightarrow 35:65, v/v). The title compound **2** was obtained as a colorless glass, $1.7 \times \text{Et}_3\text{N}$ salt, (9.6 μmol , 6.86 mg, 66%). ^1H NMR (500MHz, D_2O) δ 8.54 (s, 1H, H-8), 8.26 (s, 1H, H-2), 6.14 (d, 1H, $J = 5.0$ Hz, H-1'), 4.79-4.76 (m, 1H, H-2', obstructed by H_2O peak), 4.56-4.52 (m, 1H, H-3'), 4.42-4.38 (m, 1H, H-4'), 4.26-4.21 (m, 3H, H-5_{a,b}, H-2''), 4.17-4.14 (m, 1H, H-3''), 4.13-4.07 (m, 1H, H-5''_a), 4.03-3.92 (m, 3H, H-1''_a, 4'', 5''_b), 3.74 (dd, 1H, $J = 8.5$ Hz, $J = 3.5$ Hz, H-1''_b), 3.10 (m, 10H, NCH_2CH_3), 1.18 (m, 15H, NCH_2CH_3). ^{31}P NMR (202MHz, D_2O) δ -11.35 (m, pyrophosphate). $^{13}\text{C}\{^1\text{H}\}$ NMR (126MHz, D_2O) δ 155.2 (C-6), 152.3 (C-2), 149.1 (C-4) HMBC, 140.0 (C-8), (C-5) not observed (HMBC), 86.8 (C-1'), 83.9 (d, $J = 7.6$ Hz, C-4'), 80.3 (d, $J = 7.6$ Hz, C-4''), 74.2 (C-2'), 72.2 (C-5''_{a,b}), 71.6 (C-3''), 70.9 (C-2''), 70.4 (C-3'), 65.5 (d, $J = 3.8$ Hz, C-5''_{a,b}), 65.2 (d, $J = 3.8$ Hz, C-5'), 46.6 (NCH_2CH_3), 8.2 (NCH_2CH_3). HRMS (ES^-) calcd for $\text{C}_{15}\text{H}_{22}\text{N}_5\text{O}_{13}\text{P}_2$ 542.0695 $[\text{M}-\text{H}]^-$, found 542.0716. UV (H_2O , pH 7.4) λ_{max} 259 nm (ϵ 15958 L/mol.cm). HPLC $t_{\text{R}} = 2.93$ min.

1''- α/β -2''-Deoxyadenosine diphosphoriboside (2''-deoxy-ADPR) (3). 2-Deoxyribose-5-phosphate tributylammonium salt **11** (8 mg, 0.029 mmol) and MgCl_2 (5.6 mg, 0.058 mmol) were dissolved in DMF (1.5 mL), co-evaporated with dry toluene (2 \times) and evaporated to dryness. Dry DMF (500 μL) was added to the solid residue and the mixture was stirred at 20 $^\circ\text{C}$ until all solids dissolved (5-10 min). Then AMP-imidazolide **30** (9 mg, 0.0215 mmol) was added dropwise to the mixture in DMF (700 μL). The mixture was stirred at 20 $^\circ\text{C}$ for 16 h (HPLC showed a complex reaction mixture comprising 3-4 products, including AMP and AMP dinucleotide). The solvent was evaporated in *vacuo* and crude product was dissolved in triethylammonium bicarbonate buffer (TEAB, 3 mL, 0.1M) and purified by semi-preparative, reverse-phase HPLC using a gradient of TEAB (0.1M)-acetonitrile (95:5 \rightarrow 35:65, v/v). The title compound was obtained as a colorless glass, as a mixture of anomers β/α 0.45:0.55, $1.7 \times \text{Et}_3\text{N}$ salt, (1.72 μmol , 1.23 mg, 8%). ^1H NMR (500MHz, D_2O) δ 8.43 (s, 1H, H-8), 8.19 (s, 1H, H-2), 6.06 (d, 1H, $J = 10.0$ Hz, H-1'), 5.49-5.46 (m, 0.45H, H-1'' β), 5.42-5.39 (m, 0.55H, H-1'' α), 4.70-

4.68 (m, 1H, H-2', obstructed by H₂O peak), 4.47-4.43 (m, 1H, H-3'), 4.37-4.33 (m, 0.45H, H-3''β), 4.33-4.29 (m, 1H, H-4'), 4.23-4.19 (m, 0.55H, H-3''α), 4.16-4.10 (m, 2.45H, H-4''α and H-5'), 3.93-3.89 (m, 1.45H, H-4''β and H-5''β), 3.87-3.83 (m, 1H, H-5''α), 3.12 (q, 10H, *J* = 7.5 Hz, NCH₂CH₃), 2.30-2.23 (m, 0.55H, H-2''a-α), 2.06-2.01 (m, 0.9H, H-2''a,b-β), 1.77-1.70 (m, 0.55H, H-2''b-α), 1.19 (t, 15H, *J* = 7.5 Hz, NCH₂CH₃). ³¹P NMR (202MHz, D₂O) δ -11.34 (m, pyrophosphate). ¹³C {¹H} NMR (126MHz, D₂O) δ 155.7 (C-6), 152.9 (C-2), 149.1 (C-4), 139.8 (C-8), 118.6 (C-5), 98.4 (C-1''β), 98.1 (C-1''α), 86.7 (C-1'), 84.0 and 83.9 (m, C-4', 4''α, 4''β), 74.2 (C-2'), 71.4 (C-3''β), 71.0 (C-3''α), 70.4 (C-3'), 66.4 (d, *J* = 3.8 Hz, C-5''β), 65.6 (d, *J* = 2.5 Hz, C-5''α), 65.2 (d, *J* = 3.8 Hz, C-5'), 46.6 (NCH₂CH₃), 40.6 (C-2''α, β), 8.3 (NCH₂CH₃). HRMS (ES⁻) calcd for C₁₅H₂₂N₅O₁₃P₂ 542.0695 [M-H]⁻, found 542.0690; calcd for C₁₅H₂₂N₅NaO₁₃P₂ 565.0587 M+Na⁺, found 565.0579. HPLC *t*_R = 3.10 min.

1''-α/β-3''-Deoxyadenosine diphosphoriboside (3''-deoxy-ADPR) (4). 3-Deoxy-D-ribofuranose-5-phosphate tributylammonium salt **12** (10 mg, 0.025 mmol) and MgCl₂ (4.9 mg, 0.05 mmol) were dissolved in DMF (1 mL), co-evaporated with dry toluene (2×) and evaporated to dryness. Dry DMF (600 μL) was added to the solid residue and the mixture was stirred at 20 °C until all solids were dissolved (5-10 min). Then AMP-imidazolide **30** (9.4 mg, 0.023 mmol) was added dropwise to the mixture in DMF (400 μL). The mixture was stirred at 20 °C for 16 h. Solvent was evaporated in *vacuo* and crude product was dissolved in triethylammonium bicarbonate buffer (TEAB, 3 mL, 0.1M) and purified by semi-preparative, reverse-phase HPLC using a gradient of TEAB (0.1M)-acetonitrile (95:5 → 35:65, v/v). The title compound **4** was obtained as a colorless glass, 1.7 × Et₃N salt, (3.88 μmol, 2.8 mg, 17%). ¹H NMR (500MHz, D₂O, water suppression exp.) δ 8.37 (s, 1H, H-8), 8.11 (s, 1H, H-2), 5.98 (d, 1H, *J* = 6.0 Hz, H-1'), 5.10 (d, 0.3H, *J* = 4.0 Hz, H-1''α), 5.06 (s, 0.7H, H-1''β), 4.63 (t, 1H, *J* = 5.5 Hz, H-2'), 4.40-4.37 (m, 1H, H-3'), 4.34-4.27 (m, 1H, H-4''α, β), 4.26-4.22 (m, 1H, H-4'), 4.12-4.04 (m, 2.3H, H-2''α, 5'_{a, b}), 4.04-4.01 (m, 0.7H, H-2''β), 3.91-3.85 (m, 0.7H, H-5''β_a), 3.81-3.71 (m, 1H, H-5''α_a, β_b), 3.69-3.63 (m, 0.3H, H-5''α_b), 3.04 (q, 10H, *J* = 7.5 Hz, NCH₂), 1.90 (ddd, 0.3H, *J* = 13.0 Hz, *J* = 7.5 Hz, *J* = 2.0 Hz, H-3''α_a), 1.84-1.78 (m, 1.7H, H-3''α_b, β_{a, b}), 1.12 (t, 15H, *J* = 7.5 Hz,

NCH₂CH₃). ³¹PNMR (202MHz, D₂O) δ -11.17 (m, α -pyrophosphate), -11.46 (m, β -pyrophosphate). ¹³C{¹H}NMR (126MHz, D₂O) δ 155.1 (C-6), 152.1 (C-2), 149.0 (C-4), 139.9 (C-8), 118.5 (C-5), 102.0 (C-1'' β), 96.5 (C-1'' α), 86.7 (C-1'), 83.9 (d, J = 8.7 Hz, C-4'), 78.2 (d, J = 8.4 Hz, C-4'' β), 75.5 (d, J = 8.4 Hz, C-4'' α), 75.3 (C-2'' β), 74.2 (C-2'), 70.3 (C-3'), 70.3 (C-2'' α), 68.8 (d, J = 5.2 Hz, C-5'' β), 67.8 (d, J = 4.8 Hz, C-5'' α), 65.1 (d, J = 5.0 Hz, C-5'), 46.5(NCH₂), 32.3 (C-3'' β), 31.5 (C-3'' α), 8.1 (NCH₂CH₃). HRMS (ES⁻) calcd for C₁₅H₂₂N₅O₁₃P₂ 542.0695, [M-H]⁻ 542.0716. HPLC t_R = 2.87 min.

2'-Deoxy AMP imidazolide: 2'-deoxy-adenosine-5'-phosphoryl imidazolide (31)

2'-Deoxy-AMP was treated in the same way as AMP to generate the mono Bu₃N salt (**29**).

2'-Deoxy-AMP mono-tributylammonium salt **29** (145 mg, 0.28 mmol) and imidazole (191 mg, 2.8 mmol) were put into the flask and co-evaporated with EtOH (2 \times) and with toluene (2 \times). AldrithiolTM (186 mg, 0.84 mmol) was added to the flask followed by dry DMF (1.5 mL) and stirred until all solid matter was dissolved. Triethylamine (156 μ L, 1.13 mmol, distilled-dry-stored over KOH in the dark) and triphenylphosphine (221 mg, 0.85 mmol) were added to the mixture and the mixture was stirred at 20 °C for 16 hours. The reaction mixture was cooled to 5 °C and a cold solution of NaI (337 mg, 2.25 mmol) in dry acetone (15 mL) was added at 5 °C. The white precipitate was filtered off and washed with cold acetone. The product was dried under high vacuum and stored under argon (crude compound, hygroscopic, yield was not quantified). ¹HNMR (400MHz, *d*₄-MeOD) δ 8.36 (s, H-8), 8.18 (s, H-2), 7.87-7.86 (m, 1H, Imidazole), 7.26-7.24 (m, 1H, Imidazole), 6.98-6.96 (m, 1H, Imidazole), 6.44 (dd, 1H, J = 7.6 Hz, J = 1.2 Hz, H-1'), 4.49 (q, 1H, J = 2.8 Hz, H-3'), 4.08-4.04 (m, 1H, H-4'), 3.99-3.95 (m, 2H, H-5'), 2.75 (ddd, 1H, J = 13.6 Hz, J = 7.6 Hz, J = 1.6 Hz, H-2'a), 2.42 (dq, 1H, J = 13.6 Hz, J = 2.8 Hz, H-2'b). ³¹PNMR (160MHz, *d*₄-MeOH) δ -9.00 (s, P). ¹³C{¹H}NMR (100MHz, *d*₄-MeOD) δ 157.3 (C-6), 153.8 (C-2), 150.5 (C-4), 140.9 (C-8), 140.7 (d, J = 5.5 Hz, Imidazole), 129.6 (d, J = 10.5 Hz, Imidazole), 121.3 (d, J = 5.7 Hz, Imidazole), 120.3 (C-5), 87.5 (d, J = 8.7 Hz, C-4'),

85.5 (C-1'), 72.9 (C-3'), 67.0 (d, $J = 5.9$ Hz, C-5'), 41.2 (C-2'). HRMS (ES^-) calcd for $\text{C}_{13}\text{H}_{15}\text{N}_7\text{O}_5\text{P}$ 380.0878 $[\text{M-H}]^-$, found 380.0883.

1'', 2'-Dideoxyadenosine diphosphoriboside (1'', 2'-deoxy ADPR) (5). 1-Deoxy-D-ribofuranose-5-phosphate tributylammonium salt **10** (12 mg, 0.03 mmol) and MgCl_2 (5.9 mg, 0.06 mmol) were dissolved in DMF (1 mL), co-evaporated with dry toluene (2 \times) and evaporated to dryness. Dry DMF (600 μL) was added to the solid residue and the suspension was stirred at 20 $^\circ\text{C}$ until all solids were dissolved (5-10 min). Then 2'-dAMP-imidazolidine **31** (10.8 mg, 0.027 mmol) was added dropwise in DMF (400 μL). The mixture was stirred at 20 $^\circ\text{C}$ for 3 h. HPLC showed consumption of all starting material. The solvent was evaporated in *vacuo* and the crude product was dissolved in triethylammonium bicarbonate buffer (TEAB, 10 mL, 0.1M) and purified by semi-preparative, reverse-phase HPLC using a gradient of TEAB (0.1M)-acetonitrile (95/5 \rightarrow 35/65, v/v). The title compound **5** was obtained as a colorless amorphous solid (11.3 μmol , 8.02 mg, 42%, $1.8 \times \text{Et}_3\text{N}$ salt). ^1H NMR (500MHz, D_2O) δ 8.47 (s, 1H, H-8), 8.23 (s, 1H, H-2), 6.50 (t, 1H, $J = 5.0$ Hz, H-1'), 4.79-4.74 (m, 1H, H-3', obstructed by water peak), 4.31-4.28 (m, 1H, H-4'), 4.25-4.21 (m, 1H, H-2''), 4.19-4.11 (m, 3H, H-3'', 5'_{a,b}), 4.10-4.05 (m, 1H, H-5''_a), 4.02-3.92 (m, 3H, H-1''_a, 4'', 5''_b), 3.74 (dd, 1H, $J = 10.0$ Hz, $J = 2.5$ Hz, H-1''_b), 3.19 (q, 10H, $J = 7.5$ Hz, NCH_2CH_3), 2.83 (quint, 1H, $J = 7.0$ Hz, H-2'_a), 2.59 (dq, 1H, $J = 14.0$ Hz, $J = 3.0$ Hz, H-2'_b), 1.27 (t, 15H, $J = 7.0$ Hz, NCH_2CH_3). ^{31}P NMR (202MHz, D_2O) δ -11.33 (m, pyrophosphate). $^{13}\text{C}\{^1\text{H}\}$ NMR (126MHz, D_2O) δ 155.5 (C-6), 152.7 (C-2), 148.7 (C-4), 139.9 (C-8), 118.6 (C-5), 85.7 (d, $J = 8.6$ Hz, C-4'), 83.6 (C-1'), 80.3 (d, $J = 8.4$ Hz, C-4''), 72.2 (C-1''), 71.5 (C-3''), 71.2 (C-3'), 70.9 (C-2''), 65.5 and 65.4 (2 \times d, $J = 5.3$ Hz, $J = 4.7$ Hz, C-5', 5''), 64.6 (NCH_2CH_3), 39.0 (C-2'), 8.2 (NCH_2CH_3). HRMS (ES^-) calcd for $\text{C}_{15}\text{H}_{22}\text{N}_5\text{O}_{12}\text{P}_2$ 526.0746 $[\text{M-H}]^-$, found 526.0768. HPLC $t_R = 3.7$ min.

1''- α/β -3'', 2'-Dideoxyadenosine diphosphoriboside (3'', 2'-dideoxy-ADPR) (6). 3-Deoxy-D-ribofuranose-5-phosphate tributylammonium salt **12** (12 mg, 0.03 mmol) and MgCl_2 (5.9 mg, 0.06 mmol) were dissolved in DMF (1 mL), co-evaporated with dry toluene (2 \times) and evaporated to dryness.

Dry DMF (600 μ L) was added to the solid residue and the mixture was stirred at 20 °C until all solids were dissolved (5-10 min). Then 2'-deoxy-AMP-imidazolide **31** (10.8 mg, 0.027 mmol) was added dropwise in DMF (400 μ L). The mixture was stirred at 20 °C for 16 h. HPLC showed consumption of all starting material. The solvent was evaporated in *vacuo* and the crude product was dissolved in triethylammonium bicarbonate buffer (TEAB, 10 mL, 0.1M) and purified by semi-preparative, reverse-phase HPLC using a gradient of TEAB (0.1M)-acetonitrile (95/5 \rightarrow 35/65, v/v). The title compound **6** was obtained as a colorless amorphous solid (7.5 μ mol, 5.32 mg, 27%, $1.8 \times \text{Et}_3\text{N}$ salt). ^1H NMR (500MHz, D_2O) δ 8.48 (s, 1H, H-8), 8.24 (s, 1H, H-2), 6.51 (t, 1H, $J = 7.0$ Hz, H-1'), 5.26 (d, 0.3H, $J = 4.0$ Hz, H-1'' α), 5.21 (s, 0.7H, H-1'' β), 4.78-4.75 (m, 1H, H-3', obstructed by water signal), 4.49-4.39 (m, 1H, H-4'' α , β), 4.31-4.23 (m, 1.3H, H-2'' α , 4'), 4.20-4.11 (m, 2.7H, H-2'' β , 5' $_{a,b}$), 4.05-3.98 (m, 0.7H, H-5'' β_a), 3.94-3.84 (m, 1H, H-5'' α_a , β_b), 3.82-3.76 (m, 0.3H, H-5'' α_b), 3.19 (q, 11H, $J = 7.5$ Hz, NCH_2), 2.84 (ddd, 1H, $J = 7.5$ Hz, $J = 6.5$ Hz, $J = 1.0$ Hz, H-2' $_a$), 2.63-2.57 (m, 1H, H-2' $_b$), 2.10-2.01 (m, 0.3H, H-3'' α_a), 2.00-1.92 (m, 1.7H, H-3'' α_b , β_a , β_b), 1.27 (t, 16.5H, $J = 7.5$ Hz, NCH_2CH_3). ^{31}P NMR (202MHz, D_2O) δ -11.12 (m, α -pyrophosphate), -11.38 (m, β -pyrophosphate). $^{13}\text{C}\{^1\text{H}\}$ NMR (126MHz, D_2O) δ 155.6 (C-6), 152.7 (C-2), 148.7 (C-4), 139.9 (C-8), 118.6 (C-5), 102.1 (C-1'' β), 97.6 (C-1'' α), 85.8 (d, $J = 8.8$ Hz, C-4'), 83.6 (C-1'), 78.3 (d, $J = 8.2$ Hz, C-4'' β), 75.5 (d, $J = 8.4$, C-4'' α), 75.4 (C-2'' β), 71.3 (C-3'), 70.4 (C-2'' α), 68.8 (d, $J = 5.4$ Hz, C-5'' β), 67.9 (d, $J = 5.5$ Hz, C-5'' α), 65.4 (d, $J = 4.9$ Hz, C-5'), 46.6 (NCH_2), 39.0 (C-2'), 32.4 (C-3' β), 31.7 (C-3' α), 8.2 (NCH_2CH_3). HRMS (ES^-) calcd for $\text{C}_{15}\text{H}_{22}\text{N}_5\text{O}_{12}\text{P}_2$ 526.0746, $[\text{M}-\text{H}]^-$ 526.0761. HPLC $t_R = 3.68$ min.

Pharmacology

Materials

ADPR was obtained from Sigma Aldrich.

Cell culture

HEK293 cells were kept at 37 °C and 5% CO₂ in complete medium (DMEM with 4.5 g/L glucose, Glutamax-I, 10% FBS, 100 units/mL penicillin, and 100 µg/mL streptomycin) at 37 °C. For the maintenance of HEK293 clones with stable expression of TRPM2 400 µg/mL G418 sulfate was added to the medium.

Transfection and generation of cell lines

Generation of the clonal HEK293 cell line with stable expression of human TRPM2 has been described previously.¹⁶ In brief, HEK293 were transfected with an expression vector encoding the full-length of human TRPM2 and EGFP (pIRES2-EGFP-TRPM2). Cells that successfully integrated the expression vector were then enriched by selection with 400 µg/mL G418 sulfate (Biochrom). Clonal cell lines were established from these cells by limiting dilution. Expression of TRPM2 was confirmed by Ca²⁺ measurement and whole cell patch clamp.

Patch Clamp measurements

The day before the experiments cells from a clonal HEK293 cell line with stable expression of human TRPM2 were seeded to 35 mm tissue culture dishes at a low density. For the whole cell patch clamp experiments the culture medium was replaced by a bath solution containing 1 mM CaCl₂, 140 mM NMDG, 5 mM KCl, 3.3 mM MgCl₂, 1 mM CaCl₂, 5 mM D-glucose, 10 mM HEPES, pH 7.4. Patch pipettes were pulled from borosilicate glass capillaries with an outer diameter of 1.5 mm and an inner diameter of 1.05 mm using a Sutter P-97 and filled with a pipette solution containing 120 mM KCl, 8 mM NaCl, 1 mM MgCl₂, 10 mM HEPES, 10 mM EGTA, and 5.6 mM CaCl₂ (resulting in 200 nM free [Ca²⁺] as calculated by CaBuf software (G. Droogmans, formerly available from <ftp://ftp.cc.kuleuven.ac.be/pub/droogmans/cabuf.zip>). These pipettes had a resistance between 1.5 and 3.5 MΩ. Data were recorded using an EPC-10 amplifier and PatchMaster software (HEKA Elektronik, Germany). Fast and slow capacity transients were compensated, series resistance was compensated by

70%. After establishing whole cell configuration cells were held at -50 mV and channel activation was followed by applying voltage ramps from -85 mV to +20 mV over 140 ms every 5 s for a total of 450 s. For further analysis the maximum outward current at +15 mV during the course of the recording was taken as a measure of channel activity. To test for agonist activity ADPR analogues were included in the pipette solution at 100 μ M. To test for antagonist activity the pipette solution contained 100 μ M ADPR and 900 μ M of the ADPR analogue under test. All experiments were performed at room temperature.

Statistical analysis

The data from patch-clamp experiments were analysed using GraphPad Prism (version 7.04, GraphPad Software Inc.) Since the distribution of currents is skewed towards higher values, data were log-transformed. Log-transformed data were tested for significant differences using One-Way-Anova followed by post-hoc testing against the respective control (buffer for agonist experiments, ADPR for antagonist experiments) using Bonferroni correction for multiple testing ($\alpha=0.05$). In the charts the horizontal bar indicates the mean of the log-transformed data.

Supporting Information ^1H -NMR, ^{13}C -NMR and ^{31}P -NMR spectra for compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>

Corresponding author information *Phone: ++44-1865-271945. Fax: ++44-1865-271853. Email: barry.potter@pharm.ox.ac.uk

Author Contributions BVLP and AHG devised the overall work area. BVLP, JMW and RF devised the focused strategy. OBB synthesised the ADPR analogues supervised by JMW. MDR carried out patch clamp experiments with RF. OBB, JMW, RF and BVLP wrote the manuscript with input from all authors.

Acknowledgements BVLP is a Wellcome Trust Senior Investigator (Grant 101010). This study was supported by the Deutsche Forschungsgemeinschaft (GU 360/16-1 and Projektnummer 335447717-SFB1328 project A01 to AHG, Projektnummer 335447717-SFB1328 project A05 to RF) and Landesforschungsförderung Hamburg (Research Group ReAd Me, project 01, to AHG). The authors thank Andreas Bauche for technical support.

References:

- (1) Perraud, A. L.; Fleig, A.; Dunn, C. A.; Bagley, L. A.; Launay, P.; Schmitz, C.; Stokes, A. J.; Zhu, Q.; Bessman, M. J.; Penner, R.; et al. ADP-Ribose Gating of the Calcium-Permeable LTRPC2 Channel Revealed by Nudix Motif Homology. *Nature* **2001**, *411* (6837), 595–599.
- (2) Perraud, A.-L.; Shen, B.; Dunn, C. A.; Rippe, K.; Smith, M. K.; Bessman, M. J.; Stoddard, B. L.; Scharenberg, A. M. NUDT9, a Member of the Nudix Hydrolase Family, Is an Evolutionarily Conserved Mitochondrial ADP-Ribose Pyrophosphatase. *J. Biol. Chem.* **2003**, *278* (3), 1794–1801.
- (3) Iordanov, I.; Mihályi, C.; Tóth, B.; Csanády, L. The Proposed Channel-Enzyme Transient Receptor Potential Melastatin 2 Does Not Possess ADP Ribose Hydrolase Activity. *Elife* **2016**, *5* (12), 7250–7257.
- (4) Fonfria, E.; Marshall, I. C. B.; Benham, C. D.; Boyfield, I.; Brown, J. D.; Hill, K.; Hughes, J. P.; Skaper, S. D.; McNulty, S. TRPM2 Channel Opening in Response to Oxidative Stress Is Dependent on Activation of Poly(ADP-Ribose) Polymerase. *Br. J. Pharmacol.* **2004**, *143* (1), 186–192.
- (5) Buelow, B.; Song, Y.; Scharenberg, A. M. The Poly(ADP-Ribose) Polymerase PARP-1 Is Required for Oxidative Stress-Induced TRPM2 Activation in Lymphocytes. *J. Biol. Chem.* **2008**, *283* (36), 24571–24583.

- (6) Zhang, W.; Hirschler-laszkiewicz, I.; Tong, Q.; Conrad, K.; Sun, S.; Penn, L.; Barber, D. L.; Stahl, R.; Carey, D. J.; Cheung, J. Y.; et al. TRPM2 Is an Ion Channel That Modulates Hematopoietic Cell Death through Activation of Caspases and PARP Cleavage. *Am. J. Physiol. Cell Physiol* **2006**, 290 (47), 1146–1159.
- (7) Yang, Y.; Jiang, G.; Zhang, P.; Fan, J. Programmed Cell Death and Its Role in Inflammation. *Mil. Med. Res.* **2015**, 2 (1), 1–12.
- (8) Fonfria, E.; Mattei, C.; Hill, K.; Brown, J. T.; Randall, A.; Benham, C. D.; Skaper, S. D.; Campbell, C. A.; Crook, B.; Murdock, P. R.; et al. TRPM2 Is Elevated in the TMCAO Stroke Model, Transcriptionally Regulated, and Functionally Expressed in C13 Microglia. *J. Recept. Signal Transduct. Res.* **2006**, 26 (3), 179–198.
- (9) Knowles, H.; Li, Y.; Perraud, A. L. The TRPM2 Ion Channel, an Oxidative Stress and Metabolic Sensor Regulating Innate Immunity and Inflammation. *Immunol. Res.* **2013**, 55 (1–3), 241–248.
- (10) Partida-Sanchez, S.; Gasser, A.; Fliegert, R.; Siebrands, C. C.; Dammermann, W.; Shi, G.; Mousseau, B. J.; Sumoza-Toledo, A.; Bhagat, H.; Walseth, T. F.; et al. Chemotaxis of Mouse Bone Marrow Neutrophils and Dendritic Cells Is Controlled by ADP-Ribose, the Major Product Generated by the CD38 Enzyme Reaction. *J. Immunol.* **2007**, 179 (11), 7827–7839.
- (11) Sumoza-Toledo, A.; Lange, I.; Cortado, H.; Bhagat, H.; Mori, Y.; Fleig, A.; Penner, R.; Partida-Sanchez, S. Dendritic Cell Maturation and Chemotaxis Is Regulated by TRPM2-Mediated Lysosomal Ca²⁺ Release. *FASEB Journal*. 2011, pp 3529–3542.
- (12) Wang, G.; Cao, L.; Liu, X.; Bai, C.; Malik, A. B.; Xu Correspondence, J.; Sieracki, N. A.; Di, A.; Wen, X.; Chen, Y.; et al. Oxidant Sensing by TRPM2 Inhibits Neutrophil Migration and Mitigates Inflammation In Brief Oxidant Sensing by TRPM2 Inhibits Neutrophil Migration and Mitigates Inflammation. *Dev. Cell* **2016**, 38, 1–10.

- (13) Yamamoto, S.; Shimizu, S.; Kiyonaka, S.; Takahashi, N.; Wajima, T.; Hara, Y.; Negoro, T.; Hiroi, T.; Kiuchi, Y.; Okada, T.; et al. TRPM2-Mediated Ca^{2+} Influx Induces Chemokine Production in Monocytes That Aggravates Inflammatory Neutrophil Infiltration. *Nat. Med.* **2008**, *14* (7), 738–747.
- (14) Wehrhahn, J.; Kraft, R.; Harteneck, C.; Hauschildt, S. Transient Receptor Potential Melastatin 2 Is Required for Lipopolysaccharide-Induced Cytokine Production in Human Monocytes. *J. Immunol.* **2010**, *184* (5), 2386–2393.
- (15) Melzer, N.; Hicking, G.; Göbel, K.; Wiendl, H. TRPM2 Cation Channels Modulate T Cell Effector Functions and Contribute to Autoimmune CNS Inflammation. *PLoS One* **2012**, *7* (10), e47617.
- (16) Gelderblom, M.; Melzer, N.; Schattling, B.; Göb, E.; Hicking, G.; Arunachalam, P.; Bittner, S.; Ufer, F.; Herrmann, A. M.; Bernreuther, C.; et al. Transient Receptor Potential Melastatin Subfamily Member 2 Cation Channel Regulates Detrimental Immune Cell Invasion in Ischemic Stroke. *Stroke* **2014**, *45* (11), 3395–3402.
- (17) Belrose, J. C.; Jackson, M. F. TRPM2: A Candidate Therapeutic Target for Treating Neurological Diseases. *Acta Pharmacol. Sin.* **2018**, *39* (5), 722–732.
- (18) Li, J.; Gao, Y.; Bao, X.; Li, F.; Yao, W.; Feng, Z.; Yin, Y. TRPM2: A Potential Drug Target to Retard Oxidative Stress. *Front. Biosci. (Landmark Ed.)* **2017**, *22*, 1427–1438.
- (19) Zhang, Z.; Tóth, B.; Szollosi, A.; Chen, J.; Csanády, L. Structure of a TRPM2 Channel in Complex with Ca^{2+} Explains Unique Gating Regulation. *Elife* **2018**, *7*, e36409.
- (20) Huang, Y.; Winkler, P. A.; Sun, W.; Lü, W.; Du, J. Architecture of the TRPM2 Channel and Its Activation Mechanism by ADP-Ribose and Calcium. *Nature* **2018**, *562* (7725), 145–149.

- (21) Wang, L.; Fu, T.-M.; Zhou, Y.; Xia, S.; Greka, A.; Wu, H. Structures and Gating Mechanism of Human TRPM2. *Science* **2018**, *362*, eaav4809.
- (22) Moreau, C.; Kirchberger, T.; Swarbrick, J. M.; Bartlett, S. J.; Fliegert, R.; Yorgan, T.; Bauche, A.; Harneit, A.; Guse, A. H.; Potter, B. V. L. Structure-Activity Relationship of Adenosine 5'-Diphosphoribose at the Transient Receptor Potential Melastatin 2 (TRPM2) Channel: Rational Design of Antagonists. *J. Med. Chem.* **2013**, *56* (24), 10079–10102.
- (23) Fliegert, R.; Bauche, A.; Wolf Pérez, A.-M.; Watt, J. M.; Rozewitz, M. D.; Winzer, R.; Janus, M.; Gu, F.; Rosche, A.; Harneit, A.; et al. 2'-Deoxyadenosine 5'-Diphosphoribose Is an Endogenous TRPM2 Superagonist. *Nat. Chem. Biol.* **2017**, *13* (9), 1036–1044.
- (24) Morgan, A. J.; Wang, Y. K.; Roberts, M. F.; Miller, S. J. Chemistry and Biology of Deoxy-myoinositol Phosphates: Stereospecificity of Substrate Interactions within an Archaeal and a Bacterial IMPase. *J. Am. Chem. Soc.* **2004**, *126*, 15370-15371.
- (25) Fowler, B. S.; Laemmerhold, K. M.; Miller, S. J. Catalytic Site-Selective Thiocarbonylations and Deoxygenations of Vancomycin Reveal Hydroxyl-Dependent Conformational Effects. *J. Am. Chem. Soc.* **2012**, *134*, 9755-9761.
- (26) Jordan, P. A.; Miller, S. J. An Approach to the Site-Selective Deoxygenation of Hydroxyl Groups Based on Catalytic Phosphoramidite Transfer. *Angew. Chem. Int. Ed.* **2012**, *51*, 2907-2911.
- (27) Palacios, D. S.; Dailey, I.; Siebert, D. M.; Wilcock, B. C.; Burke, M. D. Synthesis-Enabled Functional Group Deletions Reveal Key Underpinnings of Amphotericin B Ion Channel and Antifungal Activities. *PNAS* **2011**, *108*, 6733-6738.
- (28) Szpilman, A. M.; Mathorpe, J. M.; Carreira, E. M. Synthesis and Biological Studies of 35-Deoxy Amphotericin B Methyl Ester. *Angew. Chem. Int. Ed.* **2008**, *47*, 4339-4342.

- (29) Fliegert, R.; Watt, J. M.; Schöbel, A.; Rozewitz, M. D.; Moreau, C.; Kirchberger, T.; Thomas, M. P.; Sick, W.; Araujo, A. C.; Harneit, A.; et al. Ligand-Induced Activation of Human TRPM2 Requires the Terminal Ribose of ADPR and Involves Arg1433 and Tyr1349. *Biochem. J.* **2017**, *474* (13), 2159–2175.
- (30) Li, W.; Niu, Y.; Xiong, D. C.; Cao, X.; Ye, X. S. Highly Substituted Cyclopentane-CMP Conjugates as Potent Sialyltransferase Inhibitors. *J. Med. Chem.* **2015**, *58* (20), 7972–7990.
- (31) Graham, S. M.; Pope, S. C. Selective Phosphitylation of the Primary Hydroxyl Group in Unprotected Carbohydrates and Nucleosides. *Org. Lett.* **1999**, *1* (5), 733–736.
- (32) Bessodes, M.; Komiotis, D.; Antonakis, K. Rapid and Selective Detritylation of Primary Alcohols Using Formic Acid. *Tetrahedron Lett.* **1986**, *27* (5), 579–580.
- (33) Skorupa, E.; Dmochowska, B.; Pellowaska-Januszek, L.; Wojnowski, W.; Chojnacki, J.; Wiśniewski, A. Synthesis and Structure of Selected Quaternary *N*-(1,4-Anhydro-5-Deoxy-2,3-*O*-Isopropylidene-*D,L*-Ribitol-5-yl)Ammonium Salts. *Carbohydr. Res.* **2004**, *339* (14), 2355–2362.
- (34) Moravcová, J.; Čapková, J.; Staněk, J. One-Pot Synthesis of 1,2-*O*-Isopropylidene- α -d-Xylofuranose. *Carbohydr. Res.* **1994**, *263* (1), 61–66.
- (35) Ko, H.; Das, A.; Carter, R. L.; Fricks, I. P.; Zhou, Y.; Ivanov, A. A.; Melman, A.; Joshi, B. V.; Kováč, P.; Hajduch, J.; et al. Molecular Recognition in the P2Y₁₄ receptor: Probing the Structurally Permissive Terminal Sugar Moiety of Uridine-5'-Diphosphoglucose. *Bioorganic Med. Chem.* **2009**, *17* (14), 5298–5311.
- (36) Dabrowski-Tumanski, P.; Kowalska, J.; Jemielity, J. Efficient and Rapid Synthesis of Nucleoside Diphosphate Sugars from Nucleoside Phosphorimidazolides. *European J. Org. Chem.* **2013**, *2013* (11), 2147–2154.

- (37) Hamill, O. P.; Marty, A.; Neher, E.; Sakmann, B.; Sigworth, F. J. Improved Patch-Clamp Techniques for High-Resolution Current Recording from Cells and Cell-Free Membrane Patches. *Pflügers Arch. Eur. J. Physiol.* **1981**, *391* (2), 85–100.
- (38) Molleman, A. Patch Clamping: An Introductory Guide to Patch Clamp Electrophysiology; John Wiley & Sons, 2003.
- (39) Tóth, B.; Iordanov, I.; Csanády, L. Putative Chanzyme Activity of TRPM2 Cation Channel Is Unrelated to Pore Gating. *Proc. Natl. Acad. Sci. U. S. A.* **2014**, *111* (47), 16949–16954.
- (40) Luo, X.; Li, M.; Zhan, K.; Yang, W.; Zhang, L.; Wang, K.; Yu, P.; Zhang, L. Selective Inhibition of TRPM2 Channel by Two Novel Synthesized ADPR Analogues. *Chem. Biol. Drug Des.* **2018**, *91* (2), 552–566.
- (41) Grubisha, O.; Rafty, L. A.; Takanishi, C. L.; Xu, X.; Tong, L.; Perraud, A. L.; Scharenberg, A. M.; Denu, J. M. Metabolite of SIR2 Reaction Modulates TRPM2 Ion Channel. *J. Biol. Chem.* **2006**, *281* (20), 14057–14065.
- (42) Jackson, M. D.; Denu, J. M. Structural Identification of 2'- and 3'-O-Acetyl-ADP-Ribose as Novel Metabolites Derived from the Sir2 Family of Beta-NAD⁺-Dependent Histone/Protein Deacetylases. *J. Biol. Chem.* **2002**, *277* (21), 18535–18544.
- (43) Neidle, S. *Principles of Nucleic Acid Structure*; Elsevier, **2008** pp 20-33.
- (44) Saenger, W. *Principles of Nucleic Acid Structure*; Springer-Verlag, **1984** pp51-104.
- (45) Kühn, F. J. P.; Watt, J. M.; Potter B. V. L.; Lückhoff, A. Different substrate specificities of the two ADPR interaction sites in TRPM2 channels of *Nematostella vectensis* and the role of IDPR. *Sci Rep.* **2019**, *9*, 4985.

